

REVIEW

Open Access



# The 30-year evolution of airway pressure release ventilation (APRV)

Sumeet V. Jain<sup>1</sup>, Michaela Kollisch-Singule<sup>1</sup>, Benjamin Sadowitz<sup>1</sup>, Luke Dombert<sup>1</sup>, Josh Satalin<sup>1\*</sup>, Penny Andrews<sup>2</sup>, Louis A. Gatto<sup>1,3</sup>, Gary F. Nieman<sup>1</sup> and Nader M. Habashi<sup>2</sup>

\* Correspondence:

satalinj@upstate.edu

<sup>1</sup>Department of Surgery, SUNY  
Upstate Medical University, 750 E  
Adams St, Syracuse, NY 13210, USA  
Full list of author information is  
available at the end of the article

## Abstract

Airway pressure release ventilation (APRV) was first described in 1987 and defined as continuous positive airway pressure (CPAP) with a brief release while allowing the patient to spontaneously breathe throughout the respiratory cycle. The current understanding of the optimal strategy to minimize ventilator-induced lung injury is to “open the lung and keep it open”. APRV should be ideal for this strategy with the prolonged CPAP duration recruiting the lung and the minimal release duration preventing lung collapse. However, APRV is inconsistently defined with significant variation in the settings used in experimental studies and in clinical practice. The goal of this review was to analyze the published literature and determine APRV efficacy as a lung-protective strategy. We reviewed all original articles in which the authors stated that APRV was used. The primary analysis was to correlate APRV settings with physiologic and clinical outcomes. Results showed that there was tremendous variation in settings that were all defined as APRV, particularly CPAP and release phase duration and the parameters used to guide these settings. Thus, it was impossible to assess efficacy of a single strategy since almost none of the APRV settings were identical. Therefore, we divided all APRV studies divided into two basic categories: (1) fixed-setting APRV (F-APRV) in which the release phase is set and left constant; and (2) personalized-APRV (P-APRV) in which the release phase is set based on changes in lung mechanics using the slope of the expiratory flow curve. Results showed that in no study was there a statistically significant worse outcome with APRV, regardless of the settings (F-APRV or P-APRV). Multiple studies demonstrated that P-APRV stabilizes alveoli and reduces the incidence of acute respiratory distress syndrome (ARDS) in clinically relevant animal models and in trauma patients. In conclusion, over the 30 years since the mode’s inception there have been no strict criteria in defining a mechanical breath as being APRV. P-APRV has shown great promise as a highly lung-protective ventilation strategy.

**Keywords:** APRV, Ventilator-induced lung injury, ARDS, Lung protection

## Review

### Introduction

Stock and Downs first defined airway pressure release ventilation (APRV) as maintenance of a continuous positive airway pressure (CPAP) that allows for spontaneous breaths without significant airway pressure fluctuation and a brief cyclic release phase for efficient ventilation (i.e., CPAP with release) [1]. However, the flexibility of this

definition has become an Achilles heel of sorts since a wide variety of APRV settings have all been used in the literature making comparison between studies impossible. Therefore, the current acronym “APRV” is a nebulous term identifying a ventilator mode without a precisely defined mechanical breath structure or ventilator settings. Variability in settings, and thus mechanical breath structure, significantly changes how the lung “sees” the breath being delivered. The first description of APRV published by Stock et al. in 1987 was, “APRV is a new way to administer simultaneously a supportive level of CPAP and assist CO<sub>2</sub> elimination” [2]. Thus, the original definition of APRV was simply CPAP with a release to eliminate CO<sub>2</sub>. CPAP with a release is a very general description of a mechanical breath and can define all of the mechanical breaths seen in Fig. 1. Indeed, all of these mechanical breaths were defined as APRV [2–5]. As can be readily seen, the biggest difference between these APRV breaths is the duration at inspiration and expiration. Since the original definition of APRV was CPAP with a release, which did not specify the duration of the CPAP or release time, all of these breaths fit the original definition. Over the 30 years since APRV was originally described, many combinations of CPAP and release times have been used and all defined as APRV.

In this review, we examine published original research in both animal and human trials in which the authors stated they utilized APRV, or a comparable but differently named ventilator mode. Our goals with this review were as follows: (1) to examine the historical evolution of APRV methodology; (2) to evaluate the efficacy of the APRV methodologies versus conventional positive pressure ventilation (CPPV); and (3) to determine if there are optimal APRV settings for maximal lung protection.

The evolution of APRV mostly revolves around modifying the CPAP and release time durations (time at expiration— $T_{Low}$ ) (Tables 1, 2, 3, and 4, %CPAP and  $T_{Low}$ ). However, the most significant evolution in APRV has been the development of the ability to personalize the expiratory duration to precisely meet the needs of the patient’s changing lung physiology. The advantage of this method is that expiratory duration is set to maintain and open and stable the lung, regardless the level of lung pathology. Since there was no consistency in the CPAP and release time duration in the published literature, we chose to separate APRV strategies into two categories: fixed- (F-APRV) and personalized (P-APRV) APRV. If the expiratory duration or release time was not adjusted by mechanical changes in the lung, regardless of duration of CPAP or release time, it was categorized as F-APRV. If the APRV strategy sets expiratory duration by changes in lung mechanics, using the slope of the expiratory flow curve, it was categorized as P-APRV (Fig. 1).

## Methodology

A PubMed search of the terms “Airway Pressure Release Ventilation”, “APRV”, “Bi-Vent”, “APRV/Biphasic” and “Bi-Level” and “Mechanical Ventilation” OR “Ventilator” was conducted alone or in combination. English language studies between the years 1987 and 2015 were included, and studies testing non-invasive ventilator strategies alone, reviews, editorials, and case studies were excluded, yielding 52 articles. Of the 60 articles, 32 were animal studies and 28 human studies (Tables 1, 2, 3, and 4). APRV methodologies were analyzed by examining the following settings: (1) pressure during inspiration/CPAP phase ( $P_{High}$ ); (2) time during inspiration/CPAP phase ( $T_{High}$ ); (3)

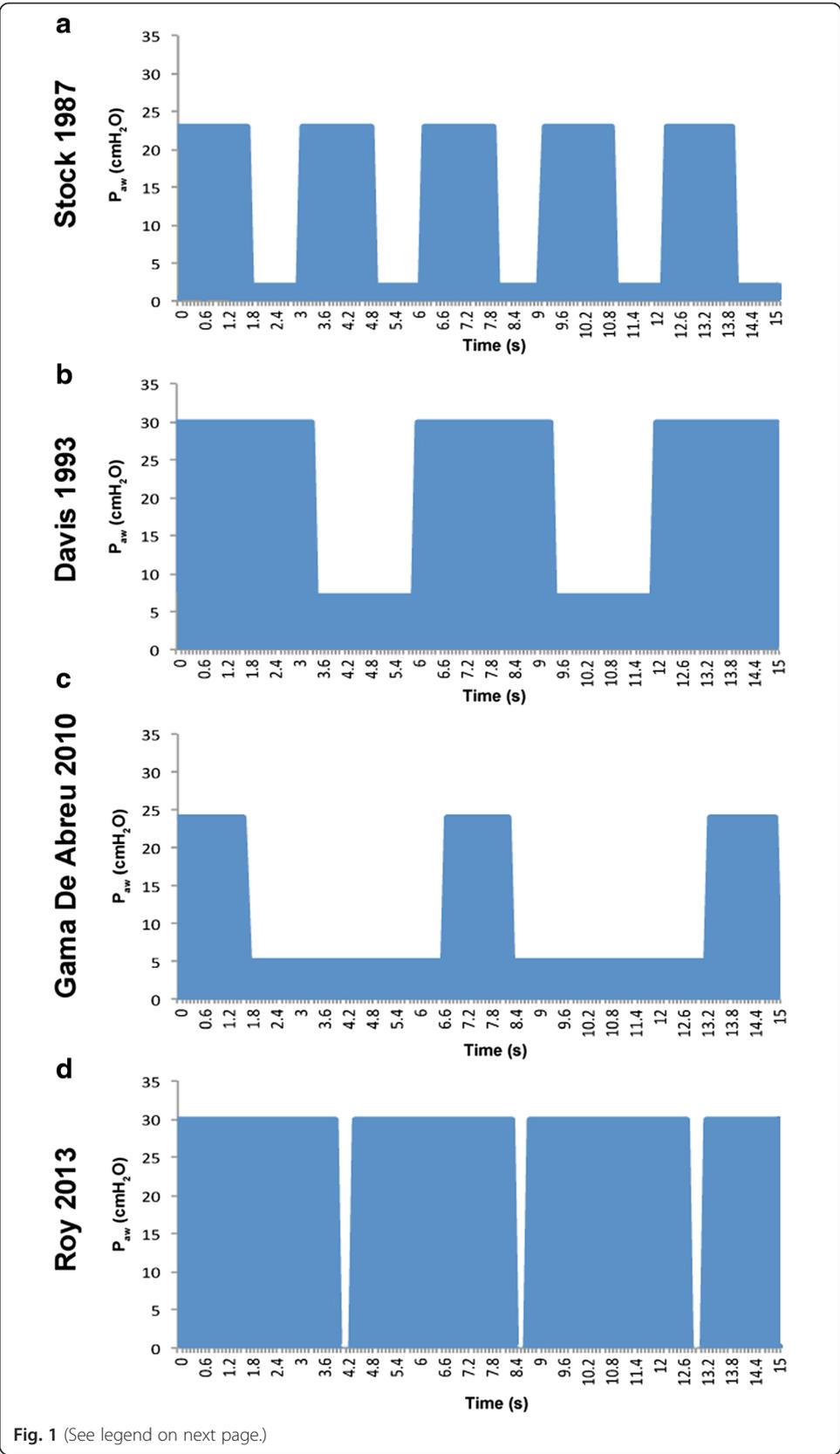


Fig. 1 (See legend on next page.)

(See figure on previous page.)

**Fig. 1** Comparison of APRV pressure waveforms. Artistic depiction of airway pressure waveforms, all of which were defined as APRV, illustrating the significant variability in what has been defined as an APRV breath. Stock in 1987 used 60 % CPAP with  $T_{Low}$  of 1.27 s and a respiratory rate (RR) of 20 [2]. Davis in 1993 used a similar %CPAP, but decreased the RR by prolonging  $T_{High}$  and  $T_{Low}$  [3]. Gama de Abreu in 2010 simulated conventional ventilation with a prolonged  $T_{Low}$  and short  $T_{High}$  [4]. Finally, Roy in 2013 used a very brief adaptive  $T_{Low}$  and large  $T_{High}$  with 90 % CPAP [5]. Of note, though the ventilator pressure is set at zero, this does not reflect true pressure as the brief  $T_{Low}$  prevents full deflation of the lung, and thus prevents end-expiratory pressure from reaching zero. Figures **a–c** are examples of fixed-APRV (F-APRV) and figure **d** of personalized APRV (P-APRV)

pressure during expiration/release phase ( $P_{Low}$ ); (4) time during expiration/release phase ( $T_{Low}$ ); and (5) percent CPAP (%CPAP) to reflect the time spent at  $P_{High}$  relative to the entire breath duration  $\left(\frac{T_{High}}{T_{High}+T_{Low}} \times 100\right)$ . In addition, we assessed the factors used to titrate these settings including respiratory rate or partial pressure of carbon dioxide ( $PCO_2$ ) for  $T_{High}$  and  $T_{Low}$  and oxygenation or plateau pressure for  $P_{High}$ .

Reviewing the evolution of APRV from its inception in 1987, we noted a major paradigm shift in the way APRV is set. Initially, the variations in settings were to the inspiratory and expiratory duration time that were fixed and not adjusted to changes in lung mechanics (Fig. 1a–c). In 2005, Habashi published a paper with a novel method of setting the expiratory duration based on changing lung mechanics identified by the slope of the expiratory flow curve (Figs. 1d and 2) [6]. This novel method of setting expiratory duration sets this personalized APRV (P-APRV) strategy (Figs. 1d and 2) apart from all other fixed setting-APRV (F-APRV) strategies.

Thus, we chose to divide our review of APRV efficacy into these two basic categories. The F-APRV breath with a relatively short inspiratory time ( $T_{High}$ ) occupying <90 % of total cycle time (Fig. 1a–c) of each breath with a fixed expiratory time ( $T_{Low}$ ) that is not adjusted based on changing lung mechanics. The second category originally described by Habashi [6] was a P-APRV breath with the following settings: (1) the inspiratory pressure ( $P_{High}$ ) is set to the desired plateau pressure; (2) the  $T_{High}$  is typically set to occupy 90 % of the total cycle time of each breath (Fig. 2a); (3) the  $T_{Low}$  set based on changes in lung mechanics by analyzing the slope of the expiratory flow curve (Fig. 2b); and (4) the expiratory pressure ( $P_{Low}$ ) is set at 0 cmH<sub>2</sub>O to minimize resistance to convective expiratory gas flow and maximize ventilation. The short duration at end expiration prevents the airway pressure from reaching atmospheric pressure, thus maintaining a positive end-expiratory pressure. Based on the above criteria, the studies reviewed were placed into either the F-APRV (Tables 1 and 3) or P-APRV (Tables 2 and 4) category.

## Results

Animal (Tables 1 and 2) and human (Tables 3 and 4) studies were examined for APRV settings and efficacy. As described above, the APRV methodologies were subdivided into two categories: F-APRV; (Tables 1 and 3) and P-APRV (Tables 2 and 4). The majority of the animal studies (69 % of total) [2, 4, 7–26] and human studies (82 % of total) [3, 27–48] were in the F-APRV category.

### How expiratory duration was personalized

The expiratory flow curve is analyzed, and the ratio of the end-expiratory flow (EEF) to the peak expiratory flow (PEF) is set so that the EEF/PEF ratio is 75 % [5, 49–62]

**Table 1** Summary of animal studies utilizing F-APRV

First author	Year	n	Animal	Study design	% CPAP	T <sub>Low</sub>	Findings
Stock [2]	1987	10	Mongrel dog	Crossover CPPV vs. APRV	58 %	1.27 s	APRV improved oxygenation with lower PIP and without cardiopulmonary compromise
Rasanen [21]	1988	10	Mongrel dog	Crossover CPPV vs. CPAP vs. APRV	50 %	1.5 s	CPPV impaired circulatory function and tissue oxygen balance, APRV had higher systemic vascular resistance and decreased pulmonary vascular resistance
Martin [17]	1991	7	Neonatal sheep	Crossover Spont vs. CPAP vs. CPPV vs. APRV	50 %	1 s	APRV augmented alveolar ventilation vs. CPAP, and had lower Paw than PPV without compromised cardiovascular function
Smith [23]	1995	5	Swine	Crossover CPAP vs. APRV	80 %	1.1 s exp flow 0	APRV maintains oxygenation without hemodynamic compromise
Neumann [19]	2001	9	Swine	Crossover CPAP vs. APRV +/- PEEP	67 %	1 s	APRV decreased O <sub>2</sub> compared with CPAP, No difference with PEEP
Hering [13]	2003	12	Swine	Crossover APRV +/- SB	50 %	N/A	APRV + SB increased oxygenation and cardiovascular function
Wrigge [24]	2003	24	Swine	Randomized prospective APRV +/- SB	50 %	1.5–2 s	APRV + SB increased oxygenation and cardiovascular function
Neumann [20]	2005	20	Swine	Randomized prospective APRV +/- SB	50 %	1.5 s	APRV + B increased ventilation in dependent lung and decreased shunt
Hering [14]	2005	12	Swine	Crossover APRV vs. SB	50 %	~1.7 s	APRV + SB improved oxygenation after lung injury
Wrigge [25]	2005	22	Swine	Randomized Prospective APRV +/- SB	50 %	1.5–2 s	APRV + SB redistributes ventilation to dependent lung regions and counters cyclic collapse
Hering [12]	2008	12	Swine	Crossover APRV +/- SB	50 %	N/A	APRV + SB improved oxygenation and splanchnic blood flow
Gama de Abreu [9]	2008	12	Swine	Crossover BiPAP + SB, PSV +/- sighs, "noisy" PSV	N/A	exp flow 0	"Noisy" CPPV improved oxygenation by redistributing perfusion
Carvalho [7]	2009	5	Swine	Crossover PSV vs. BiPAP + SB	Titrated by P <sub>aw</sub>	N/A	

**Table 1** Summary of animal studies utilizing F-APRV (Continued)

Gama de Abreu [4]	2010	10	Swine	Crossover PSV vs. BiPAP + SB	25 %	N/A	BIPAP + SB and pressure support had similar oxygenation improvement and did not improve aeration of dependent lung BIPAP + SB had lower tidal volume with comparable oxygenation and ventilation distribution
Henzler [11]	2010	20	Swine	Randomized prospective APRV +/- SB	42 %	~1.2 s	Elevated IAH impaired respiratory mechanics regardless of SB
Kreyer [16]	2010	12	Swine	Randomized Prospective APRV +/- SB	50 %	1.5–2 s exp flow 0	APRV + SB improved systemic blood flow and cerebrospinal blood flow
Matsuzawa [18]	2010	21	Rabbit	Randomized prospective CPPV vs. LTV vs. APRV	95 %	0.15 s	APRV reduced HMGB1 levels and lung water
Slim [22]	2011	7	Swine	Case series APRV	80 %	N/A	Increased $P_{aw}$ increased pulmonary capillary wedge pressure and left atrial pressure, but these may not correlate with end diastolic volume
Xia [26]	2011	24	Rabbit	Randomized prospective APRV +/- SB	50 %	N/A	APRV + SB improved oxygenation and attenuated VILI
Carvalho [8]	2014	36	Swine	Randomized prospective APRV +/- SB	50 %	N/A	APRV + SB improved oxygenation and reduced lung injury
Guldner [10]	2014	12	Swine	Crossover APRV +/- SB	50 %	~1 s	Higher levels of SB reduce global lung stress and strain with minimal changes in perfusion
Kill [15]	2014	24	Swine	Randomized prospective CPPV vs. Bilevel vs. Compression synchronized ventilation	40 %	3.6 s	CPPV and Bilevel usable during CPR, though compression synchronized ventilation was best

Number of studies: 22

$T_{Low}$  time at low pressure, CPPV conventional positive pressure ventilation, LTV low tidal volume ventilation, CPAP continuous positive airway pressure, SB spontaneous breathing, PEEP positive end-expiratory pressure, PIP peak inspiratory pressure,  $P_{aw}$  airway pressure, PSV pressure support ventilation, BiPAP biphasic positive airway pressure

**Table 2** Summary of animal studies utilizing P-APRV

First author	Year	<i>n</i>	Animal	Study design	%CPAP	$T_{Low}$	Findings
Albert [49]	2011	22	Swine	Randomized prospective, CPPV vs. LTV vs. APRV vs. HFOV	90 %	50–75 % PEF	APRV increased oxygenation and ventilation with reduced cytokines compared to LTV
Roy [57]	2012	8	Swine	Randomized prospective, CPPV vs. APRV	90 %	75 % PEF	Early APRV prevented ARDS with improved oxygenation, histopathology, and surfactant protein preservation
Emr [62]	2013	16	Rat	Randomized prospective, spont vs. CPPV vs. APRV	90 %	75 % PEF	Early APRV prevented ARDS with improved oxygenation and histopathology
Roy [5]	2013	12	Swine	Randomized prospective, sham vs. LTV vs. APRV	90 %	75 % PEF	Early APRV prevented ARDS with improved oxygenation and histopathology with reduced inflammatory markers
Roy [61]	2013	9	Rat	Randomized prospective, CPPV vs. APRV	90 %	75 % PEF	Early APRV prevented ARDS with improved oxygenation and histopathology
Kollisch-Singule [55]	2014	8	Rat	Randomized prospective, CPPV vs. APRV	90 %	10 or 75 % PEF	APRV with low expiratory time reduced conducting airway microstrain
Kollisch-Singule [54]	2014	6	Rat	Randomized prospective, CPPV with PEEP vs. APRV	90 %	10 or 75 % PEF	APRV with low expiratory time reduced alveolar microstrain
Davies [52]	2015	22	Swine	Randomized prospective, LTV vs. APRV	90 %	75 % PEF	APRV increased oxygenation compared with LTV, APRV had a "trend towards" increased cerebral ischemia.
Arrindell [51]	2015	19	Preterm swine	Randomized prospective, CPPV vs. APRV	N/A	75 % PEF	APRV increased oxygenation without change in histopathology or inflammatory markers
Kollisch-Singule [56]	2015	14	Rat	Randomized prospective, uninjured vs. LTV vs. APRV	90 %	75 % PEF	APRV approximated control lungs best with increased homogeneity compared to LTV. LTV with high PEEP reduced heterogeneity.

Number of studies: 10

$T_{Low}$  time at low pressure, CPPV conventional positive pressure ventilation, LTV low tidal volume ventilation, CPAP continuous positive airway pressure, PEF peak expiratory flow, SB spontaneous breathing, PEEP positive end-expiratory pressure, PIP peak inspiratory pressure

**Table 3** Summary of human trials using F-APRV

First author	Year	n	Study design	%CPAP	T <sub>Low</sub>	Findings
Garner [31]	1988	14	Crossover CPPV baseline with APRV wean	N/A	1.5 s	APRV maintained similar oxygenation with >50 % reduced PIP
Rasanen [40]	1991	50	Crossover PEEP titrated CPPV vs. APRV	50 %	1.5 s	APRV maintained similar oxygenation with >50 % reduced PIP
Cane [28]	1991	18	Crossover CPPV vs. APRV	67 %	1.5 s	APRV maintained similar oxygenation and cardiopulmonary function with reduced PIP
Davis [3]	1993	15	Crossover CPPV vs. APRV	32 %	2.6 ± 0.6	APRV maintained similar oxygenation with >50 % reduced PIP and reduced PEEP
Chiang [29]	1994	18	Crossover CPPV vs. APRV	66 %	1.5 s	APRV maintained similar oxygenation with >50 % reduced PIP
Sydow [42]	1994	18	Crossover CPPV vs. APRV	80 %	0.5–0.7	APRV maintained similar oxygenation with decreased A-a gradient after 8 h and reduced PIP
Bratzke [27]	1998	20	Crossover CPPV vs. APRV	88 %	1	APRV maintained similar oxygenation with reduced PIP
Kaplan [36]	2001	12	Crossover Inverse ratio PPV vs. APRV	85 %	0.8	APRV is safe, decreases PIP and need for sedation/paralytics/pressors, increases CI
Putensen [39]	2001	30	Randomized prospective CPPV vs. APRV	Identical to CPPV	Exp flow 0	APRV + SB maintained increased oxygenation, CI, and pulmonary compliance with reduced ALI/ARDS incidence and sedative requirements
Schultz [41]	2001	15	Crossover CPPV vs. APRV	N/A	N/A	APRV maintained similar oxygenation with >50 % reduced PIP
Wrigge [45]	2001	14	Randomized prospective APRV +/- automatic tube compensation	N/A	N/A	APRV with tube compensation increased end-expiratory lung volume and minute ventilation without affecting oxygenation or cardiopulmonary status
Hering [34]	2002	12	Crossover APRV +/- SB	N/A	N/A	APRV + SB had increased renal blood flow and glomerular filtration rate
Varpula [43]	2003	33	Randomized prospective CPPV vs. APRV	N/A	Exp flow 0	APRV feasible in prone positioning and increased oxygenation at 24 h
Varpula [44]	2004	58	Randomized prospective CPPV vs. APRV	80 %	1	APRV had similar mortality and ventilator free days
Dart [30]	2005	46	Crossover CPPV vs. APRV	N/A	40–50 % PEF	APRV reduced PIP and increased oxygenation
Liu [37]	2009	58	Retrospective case-control CPPV vs. APRV	67 %	~1.5	APRV reduced pressor use/A-a gradient and increased oxygenation
Kamath [35]	2010	11	Retrospective cohort CPPV vs. APRV	70 %	1.2 ± 0.9	APRV had no adverse effects on blood pressure or urine output
Gonzalez [32]	2010	468	Case matched retrospective CPPV vs. APRV	70 %	N/A	APRV maintained similar oxygenation with reduced

**Table 3** Summary of human trials using F-APRV (Continued)

Maxwell [38]	2010	63	Randomized prospective LTV vs. APRV	N/A	25–75 % PEF	PIP and associated increased tracheostomy rate APRV had similar physiological parameters despite increased disease severity at baseline
Hanna [33]	2011	45	Retrospective case series CPPV vs. APRV	N/A	N/A	APRV had increased P/F Ratio, lung procurement rate with similar graft survival rate
Maung [46]	2012	38	Retrospective case series APRV	85 %	0.8–1	Switching from CPPV to APRV improved oxygenation and decreased PCO <sub>2</sub> without hemodynamic compromise
Maung [47]	2012	362	Retrospective case series CPPV vs. APRV	N/A	N/A	APRV had increased ventilator days.
Testerman [48]	2013	48	Case-matched retrospective APRV; obese vs. nonobese	N/A	N/A	APRV in morbidly obese similar to nonobese, though morbidly obese required extended care after discharge more often

Number of studies: 23

*T<sub>Low</sub>* time at low pressure, CPPV conventional positive pressure ventilation, LTV low tidal volume ventilation, CPAP continuous positive airway pressure, PEF peak expiratory flow, SB spontaneous breathing, PEEP positive end-expiratory pressure, PIP peak inspiratory pressure

(Fig. 2b) based on the methodology described by Habashi [6] (Tables 2 and 4). The *T<sub>Low</sub>* is thus personalized based on alterations in lung mechanics, identified by changes in the slope of the expiratory flow curve (Fig. 2b). Using EEF/PEF ratio of 75 % results in a brief *T<sub>Low</sub>* ranging from 0.3 to 0.6 s (Fig. 1d). However, recent animal experiments have shown the optimal EEF/PEF ratio necessary to open and stabilize the lung is 75 % [54–56].

**F-APRV: inspiratory duration**

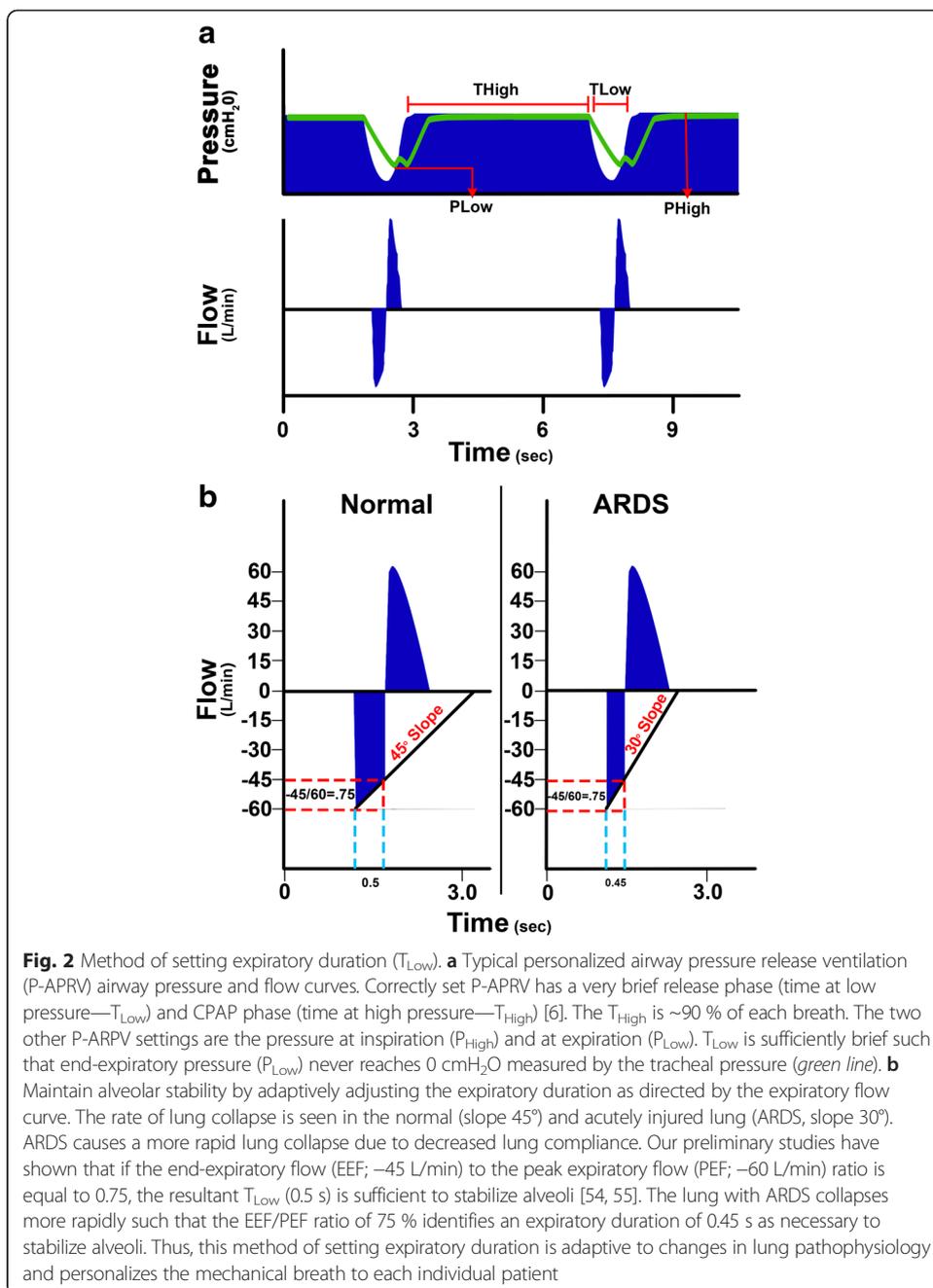
There was considerable variability in the F-APRV settings (i.e., duration at inspiration and expiration) in the papers reviewed, and in multiple papers, the settings were not defined, and thus not included in this review. Swine, dogs, and rabbits

**Table 4** Summary of human trial utilizing P-APRV

First author	Year	<i>n</i>	Study design	%CPAP	<i>T<sub>Low</sub></i>	Findings
Yoshida [60]	2009	18	Retrospective case-control LTV vs. APRV	N/A	50–75 % PEF	APRV + SB had increased oxygenation and MAP and decreased atelectasis
Walsh [58]	2011	20	Retrospective case series CPPV vs. APRV	>80 %	50–75 % PEF	APRV improved pulmonary blood flow after tetralogy of fallot repair or cavopulmonary shunt
Andrews [50]	2013	66,099	Retrospective review CPPV vs. early APRV	90 %	75 % PEF	Early APRV decreased ARDS incidence tenfold and mortality threefold
Kawaguchi [53]	2014	13	Retrospective case series CPPV vs. APRV	90 %	50–75 % PEF	APRV safe in pediatric ARDS without hemodynamic compromise
Yehya [59]	2014	104	Retrospective cohort HFOV vs. APRV	N/A	50–75 % PEF	APRV had no mortality effect compared to oscillatory ventilation as rescue treatments

Number of studies: 5

*T<sub>Low</sub>* time at low pressure, CPPV conventional positive pressure ventilation, LTV low tidal volume ventilation, PEF peak expiratory flow, SB spontaneous breathing



were used in the animals studies with the majority of studies using swine (Table 1). In only two of the animal studies were the %CPAP set at  $\leq 80$  % of the total cycle time (Table 1) [22, 23]. In one study, %CPAP was set very short at 25 % of the breath cycle time [5]. When both the animal and human studies were analyzed 23/45 studies set %CPAP  $\leq 67$  % of the breath and 13/45 had an indeterminate %CPAP (Tables 1 and 3).

#### Expiratory duration

In addition, the  $T_{Low}$  in both animal and human studies was prolonged to levels seen in conventional mechanical ventilation (Fig. 1) [3, 9, 15, 16, 24, 25, 43]. Aside from two

studies [30, 38], the  $T_{Low}$  remained fixed or was titrated based on the  $PCO_2$  as opposed to being adjusted based on changes in lung mechanics using the expiratory flow curve (Fig. 2). The  $P_{High}$  was titrated very differently in the studies reviewed, with a range between 10 cmH<sub>2</sub>O [2, 21, 28, 29, 31] and 35 cmH<sub>2</sub>O [3, 37] depending on whether the target was maximal oxygenation, maximal lung recruitment, or a specific tidal volume. In addition, most of the studies in the F-APRV group set a positive  $P_{Low} > 0$  cmH<sub>2</sub>O [2, 4, 7–11, 13, 14, 16, 18–23, 26, 40].

### **Outcome**

The majority of the studies in the F-APRV group were crossover experiments, representing 59 % of the animal studies (Table 1) and 48 % of the human trials (Table 3) [2, 4, 7, 9, 10, 12–14, 17, 19, 21, 23, 32], with the primary goal to demonstrate that APRV could be safely used without adverse effects on lung function or hemodynamic characteristics. Many crossover trials showed similar or increased oxygenation with lower peak pressures and no negative effect on hemodynamics with APRV as compared with CPPV [2, 3, 17, 27–32, 39–42], and some studies showed hemodynamic improvement with APRV [13, 24, 36, 39]. Prospective randomized trials comparing APRV and CPPV showed that APRV is safe and potentially beneficial (Table 3). In none of the studies included in this review did APRV cause a significant negative impact on the physiologic process being studied, and in some prospective randomized studies, APRV was shown beneficial (Tables 1 and 3).

### **P-APRV**

Similar to the F-APRV studies, P-APRV either resulted in improved outcomes or no change as compared with CPPV and in none of the studies was APRV shown to be harmful. Unlike the F-APRV studies that were mainly crossover studies, all of the P-APRV animal studies were randomized prospective cohort trials that compare non-protective CPPV or low tidal volume (LTV) with APRV [5, 49, 52, 54, 55, 57, 61, 62]. P-APRV was shown to reduce heterogeneity and both alveolar and alveolar duct micro-strain (i.e., change in alveolar size with applied stress) [54–56] suggesting a mechanism for the improvement in the efficacy experiments (Table 3) [5, 49, 57, 61, 62].

### **Discussion**

There has been no consensus on what parameters are essential to define a mechanical breath as being APRV, and thus, APRV settings have been inconsistent over the three decades since it was first described. However, no studies have shown that APRV is harmful or significantly inferior as compared with conventional mechanical ventilation. In 2005, Habashi clearly defined the settings for what we have termed personalized APRV or P-APRV [6]. P-APRV has a prolonged  $T_{High}$  and very brief  $T_{Low}$  duration, which is set by lung mechanics using the change in the slope of the expiratory flow curve [6]. Not only is the  $T_{Low}$  very brief in P-APRV, it is set by analyzing the slope of the expiratory flow curve and therefore adaptive to changes in the patient's lung mechanics (Fig. 2b). Using these precisely controlled APRV settings, a number of studies have shown that P-APRV recruits and stabilizes the alveoli and alveolar ducts [54–56] and reduces the incidence of ARDS in multiple animal models [5, 57, 62]. A meta-analysis has shown a reduction in ARDS incidence in trauma patients [53]. Given this

lack of consensus on how APRV should be set, the remainder of the “Discussion” will be divided into the results obtained from the F-APRV and P-APRV subgroups, rather than discuss the findings from each experiment with a different %CPAP,  $T_{Low}$ , and/or  $P_{High}$  settings.

#### **F-APRV studies**

Since, as previously mentioned, most of the animal and human studies were crossover experiments (i.e., switching from CPPV or spontaneous breathing to F-APRV in the same animal or patient), the majority of published APRV studies do not address efficacy (i.e., is APRV superior to conventional ventilation). It is important to note that APRV did not cause a significant negative impact, as compared with CPPV or spontaneous breathing, on the physiologic parameters that were measured in any of the crossover studies (Table 1 and 3). In the randomized prospective animal studies, F-APRV was shown to be beneficial. It was shown that APRV reduced lung water and HMGB1 in rabbits [18], improved systemic and cerebrospinal blood flow in swine [16], improved oxygenation and attenuated ventilator-induced lung injury (VILI) in rabbits [26], and improved oxygenation and reduced lung injury in swine [7].

In humans, Putsenen et al. showed that APRV with spontaneous breathing increased oxygenation, cardiac index, and pulmonary compliance, with reduced sedative requirements compared with CPPV in humans [39]. Varpula et al. reported similar mortality and ventilator-free days for APRV and CPPV, and also demonstrated that it is feasible to utilize APRV with prone positioning. In that study,  $T_{Low}$  was set specifically to allow expiratory flow to reach zero, which would allow the lung to collapse during expiration [43, 44]. Maxwell et al. showed no difference between low tidal volume ventilation (LTV) and APRV regarding mortality, ventilator days, ICU length of stay, or complication rates despite increased baseline disease severity in the APRV group [38]. In human retrospective trials, the APRV methodology used was not consistent among the studies analyzed (Table 3). Despite these inconsistencies, Gonzalez et al. showed that APRV reduced peak inspiratory pressure (PIP) while maintaining similar oxygenation levels [32]. Hanna et al. showed increased  $PaO_2/FiO_2$  (P/F) ratio, lung procurement rate, and graft survival when used in organ donors where the lungs were transplanted [33].

A retrospective study examining trauma patients by Maung et al. demonstrated increased ventilator days on APRV compared with an unspecified method of CPPV [47]. However, this study excluded all mortality in the interest of investigating weaning and the baseline characteristics of the groups were different, with more severe chest trauma and decreased P/F ratio at the start in the APRV group. Further, this study used respiratory therapist-guided protocol weaning only in the CPPV group, whereas the APRV group had random, non-protocolized weaning based on physician guidance.

#### **P-APRV**

Despite defining the role of tidal volume ( $V_t$ ) and plateau pressure on VILI in ARDS patients [63], our current understanding of the parameters comprising the mechanical breath that either propagate or impede progressive acute lung injury (ALI) is incomplete. The mechanism of VILI is believed to be the evolution from a normal homogeneously ventilated lung into a heterogeneously ventilated lung, with collapse and

edema-filled alveoli adjacent to open alveoli. This heterogeneity results in stress concentrators and recurrent alveolar collapse and reopening with each tidal breath that amplify lung tissue injury, instigated by the initial insult such as sepsis, trauma, or pneumonia [64–66]. Thus, the ventilation strategy that restores or maintains homogeneity would minimize VILI and obstruct progressive ALI [64, 66, 67].

P-APRV uses a 90 % CPAP phase that recruits alveoli resulting in homogeneous lung inflation (Fig. 2a) and a brief release phase with the  $T_{Low}$  personalized to the mechanics of the lung (Fig. 2b) producing a nearly static ventilated lung, which prevents alveolar collapse and reopening, thereby reducing dynamic tissue strain [54, 56]. The question is, do these APRV settings that stabilize alveoli protect the lung?

In prospective outcome animal experiments, P-APRV resulted in an increase in oxygenation along with a decrease in histopathologic injury as compared with CPPV and LTV ventilation strategies [5, 52, 57, 61, 62]. Several studies showed that P-APRV resulted in an increase in surfactant protein concentration [5, 57, 62], while two showed a reduction of inflammatory markers (Table 2) [49, 57].

Kollisch-Singule et al. [54–56] conducted three micro-anatomic studies (i.e., alveoli and alveolar ducts) that demonstrated reduced alveolar and conducting airway micro-strain as well as increased alveolar homogeneity using P-APRV in which the  $T_{Low}$  was set to maintain an EEF/PEF ratio 75 or 10 %. The  $T_{Low}$  set using an EEF/PEF ratio of 75 % was sufficiently short to stabilize alveoli and prevent alveolar collapse, whereas extending the  $T_{Low}$  (EEF/PEF ratio 10 %) resulted in alveolar collapse and instability. These studies add mechanistic support to efficacy studies showing that preemptive P-APRV reduced ARDS incidence in a clinically applicable porcine ARDS model [5, 57].

Davies et al. [52] showed increased oxygenation with P-APRV as compared with LTV, with no significant differences in cerebral ischemia in a swine model of concomitant brain and lung injury (Table 2). In patients, Yoshida et al. [60] demonstrated increased oxygenation and mean arterial pressure (MAP) with decreased atelectasis using P-APRV as compared to LTV ventilation. Walsh et al. [58] showed improved pulmonary blood flow when using P-APRV versus CPPV ventilation post-operatively after tetralogy of Fallot repair or cavopulmonary shunt in neonates (Table 4).

Furthermore, Andrews et al. [50] in a meta-analysis showed a tenfold decrease in ARDS incidence as well as a threefold decrease in mortality when compared to trauma patients with similar injuries that were treated with standard of care ventilation in 15 trauma intensive care units (Table 4). However, all of the human trials testing the P-APRV method [50, 53, 58–60] are retrospective studies, and as such, it is not clear that the precise P-APRV settings were followed, aside from the study by Andrews et al. [50] that strictly adhered to the P-APRV protocol throughout the study in trauma patients. Despite this, the human results are in concordance with multiple animal studies [5, 49, 54–57, 61, 62] that support the clinical data [50] that P-APRV may be used to reduce the incidence of acute respiratory distress syndrome (ARDS) more effectively than conventional LTV ventilation applied early or after the onset of ARDS.

#### **A note on mechanical ventilators**

An often under-analyzed aspect of mechanical breath delivery is the ventilator used to deliver the breath. Every ventilator has a specific design with differing resistances built

into the ventilator gas path, responses to spontaneous ventilation, as well as software quirks that affect breath delivery. APRV may be named in various ways based on the ventilator in use such as (1) APRV (Dräger Evita, Savina and V series, Hamilton G5), (2) Bi-Vent (Maquet Servo-i), (3) BiLevel (Engström Carestation, Puritan Bennett 840 & 980), and (4) APRV/Biphasic (Viasys Avea). However, even more problematic than the different names is the wide variation in their implementation of APRV.

In particular, the ability to control key APRV parameters such as the  $T_{Low}$  varies, and fine control of this parameter is critical to properly set P-APRV (Fig. 2b). APRV strategies on some ventilators have deviated from the original concept of a timed CPAP with a release phase, by adding pressure support (PS). This requires a trigger that creates a need for synchronization with the ventilator resulting in automated changes to both the inspiratory and expiratory duration. More specifically, the  $T_{Low}$ , which is critical to control end-expiratory lung volume and prevent airway closure, spontaneously adjusts when PS is added to this mode on these ventilators, regardless of the  $T_{Low}$  setting, producing large and variable tidal volumes leading to lung volume loss and alveolar instability. So, even if APRV was set accurately by the clinician, the ventilator may automatically adjust the duration of the release phase ( $T_{Low}$ ) ultimately allowing the lung to collapse. This effect would be similar to conventional ventilation automatically adjusting  $V_t$  or PEEP without physician input.

Furthermore, many researchers define APRV as “extreme” inverse-ratio pressure-control ventilation (IR-PCV). However, unlike IR-PCV, P-APRV does not set a defined I:E ratio. Rather, the CPAP phase is briefly interrupted by a “release phase” (i.e.,  $T_{Low}$ ) that is established by independently setting a  $T_{Low}$  based upon analysis of the expiratory flow curve. Further, patients can breathe spontaneously throughout the entire respiratory cycle in APRV, superseding the set I:E ratio. Unfortunately, many times the exact make and model of the ventilator was not reported in the papers reviewed, and thus, we could not categorize studies by the ventilator used, but it is an important factor to consider if the ventilator being used can accurately deliver P-APRV.

#### ***Current state of APRV and clinical implications***

As described previously, there is a paucity of data testing APRV with identical settings, especially in clinical trials almost all of which were retrospective or crossover studies. Rose et al. [68] reviewed the literature in 2008 and concluded that there is a lack of consistency in APRV settings making comparison with conventional ventilator strategies difficult. Another recent review by Facchin et al. [69] examined the current literature for both APRV and high-frequency oscillatory ventilation (HFOV) for treatment and prevention of ARDS; the authors concluded that there is inconsistent evidence and a lack of high quality trials to make conclusions regarding APRV or HFOV efficacy.

It is clear from the studies examined in this review that APRV has evolved from a mechanical breath defined as a prolonged CPAP phase with a brief release phase [1] into a highly sophisticated, dynamic mechanical breath with precise settings that are responsive to changes in lung physiology (i.e., P-APRV) [6]. Although the mechanical breath used by both Downs and Habashi are referred to as APRV, the actual mechanical breath created by each is very different (Fig. 1a, d) [2, 57] as is the breath created and studied in other experiments [3, 9] (Fig. 1b, c). The current use of the APRV

acronym remains an imprecise term that demands a specific definition of parameter settings to be utilized consistently. Obviously, some APRV settings are going to superior to others in their ability to protect the lung.

P-APRV appears to be an exciting and novel open lung strategy that may significantly reduce ARDS incidence, morbidity, and mortality of established ARDS. Data suggests that rather than overdistending alveoli, the extended  $T_{\text{High}}/P_{\text{High}}$  redistributes gas from the alveolar ducts to the alveoli, where it belongs [54, 55] and changes heterogeneous to homogeneous alveolar ventilation [56]. The calculated strain on alveoli was significantly reduced demonstrating that mechanical stretch on the alveolar wall was decreased. These data have led us to conclude that the extended inspiratory duration has a powerful positive impact on reducing strain at the alveolar level. We hypothesize that the mechanism of this protection (i.e., reduce alveolar strain) is secondary to increased lung volume and the number of recruited alveoli. The extended  $T_{\text{High}}$  “nudges” open alveoli over a several hours in a non-pathologic manner resulting in open lung ventilation. Since the lung becomes fully recruited with significantly improved compliance, even with relatively high tidal volumes (10–14 cm<sup>3</sup>/kg), the driving pressure is not increased (unpublished observations).

## Conclusions

Although many settings have been used, none of the studies reviewed showed a worse outcome using APRV as compared with CPPV, with many studies showing significant benefits in cardiopulmonary variables. The evolution of APRV methodology has been drastic from Downs and Stock (F-APRV) [1] to the P-APRV method of Habashi [6]. P-APRV allows for a personalized control of lung stability on a breath-to-breath basis that is not possible with other modes of ventilation. P-APRV is an adaptive, flow directed, duration dependent ventilation strategy that adapts the setting to each patient regardless of their lung pathophysiology. This personalized, adaptive mechanical breath may prove more efficacious at treating and preventing ARDS than the current standard of care. Ultimately, more studies are needed using consistent and well-defined settings to identify the optimal APRV breath necessary to maximize lung protection.

## Abbreviations

ALI: acute lung injury; APRV: airway pressure release ventilation; ARDS: acute respiratory distress syndrome; BiPAP: biphasic positive airway pressure; CPAP: continuous positive airway pressure; CPPV: conventional positive pressure ventilation; F-APRV: fixed-setting airway pressure release ventilation; HFOV: high frequency oscillatory ventilation; LTV: low tidal volume; MAP: mean arterial pressure; P/F: PaO<sub>2</sub>/FiO<sub>2</sub>; P-APRV: personalized airway pressure release ventilation; P<sub>aw</sub>: airway pressure; PCO<sub>2</sub>: partial pressure of carbon dioxide; PEEP: positive end-expiratory pressure; PEF: peak expiratory flow; P<sub>High</sub>: pressure during CPAP phase; PIP: peak inspiratory pressure; P<sub>Low</sub>: pressure during release phase; PS: pressure support; PSV: pressure support ventilation; SB: spontaneous breathing; T<sub>High</sub>: time at CPAP/P<sub>High</sub>; T<sub>Low</sub>: time at release/P<sub>Low</sub>; VILI: ventilator-induced lung injury; Vt: tidal volume.

## Authors' contributions

SVJ, MKS, BS, GFN, PA, and NMH contributed to the conception and design of this review. SVJ, LD, PA, and NMH contributed to data acquisition. SVJ, JS, GFN, and NMH helped draft the manuscript. All authors critically reviewed, read, and approved the final manuscript.

## Competing interests

GFN/NMH/PLA have presented and received honoraria and travel reimbursement at events sponsored by DrägerMedical outside of the submitted work.

GFN/NMH/PLA/SJ have lectured for Intensive Care Online (ICON).

PLA is employed by ICON.

NMH hold patents and is the founder of ICON.

Industry had no role in the design and conduct of the study; the collection, management, analysis, or interpretation of the data; or preparation, review, or approval of the manuscript.

**Author details**

<sup>1</sup>Department of Surgery, SUNY Upstate Medical University, 750 E Adams St, Syracuse, NY 13210, USA. <sup>2</sup>Multi-trauma Critical Care, R Adams Cowley Shock Trauma Center, University of Maryland Medical Center, 22 South Greene Street, Baltimore, MD, USA. <sup>3</sup>Department of Biological Sciences, 10 SUNY Cortland, Cortland, NY 13045, USA.

Received: 21 January 2016 Accepted: 3 May 2016

Published online: 20 May 2016

**References**

- Downs JB, Stock MC (1987) Airway pressure release ventilation: a new concept in ventilatory support. *Crit Care Med* 15:459–461
- Stock MC, Downs JB, Frolicher DA (1987) Airway pressure release ventilation. *Crit Care Med* 15:462–466
- Davis K Jr, Johnson DJ, Branson RD, Campbell RS, Johannigman JA, Porembka D (1993) Airway pressure release ventilation. *Arch Surg* 128:1348–1352
- Gama de Abreu M, Cuevas M, Spieth PM, Carvalho AR, Hietschold V, Stroszczyński C, Wiedemann B, Koch T, Pelosi P, Koch E (2010) Regional lung aeration and ventilation during pressure support and biphasic positive airway pressure ventilation in experimental lung injury. *Crit Care* 14:R34
- Roy S, Habashi N, Sadowitz B, Andrews P, Ge L, Wang G, Roy P, Ghosh A, Kuhn M, Satalin J, Gatto LA, Lin X, Dean DA, Vodovotz Y, Nieman G (2013) Early airway pressure release ventilation prevents ARDS—a novel preventive approach to lung injury. *Shock* 39:28–38
- Habashi NM (2005) Other approaches to open-lung ventilation: airway pressure release ventilation. *Crit Care Med* 33:S228–S240
- Carvalho AR, Spieth PM, Pelosi P, Beda A, Lopes AJ, Neykova B, Heller AR, Koch T, Gama de Abreu M (2009) Pressure support ventilation and biphasic positive airway pressure improve oxygenation by redistribution of pulmonary blood flow. *Anesth Analg* 109:856–865
- Carvalho NC, Guldner A, Beda A, Rentzsch I, Uhlig C, Dittrich S, Spieth PM, Wiedemann B, Kasper M, Koch T, Richter T, Rocco PR, Pelosi P, de Abreu MG (2014) Higher levels of spontaneous breathing reduce lung injury in experimental moderate acute respiratory distress syndrome. *Crit Care Med* 42:e702–e715
- Gama de Abreu M, Spieth PM, Pelosi P, Carvalho AR, Walter C, Schreiber-Ferstl A, Aikele P, Neykova B, Hubler M, Koch T (2008) Noisy pressure support ventilation: a pilot study on a new assisted ventilation mode in experimental lung injury. *Crit Care Med* 36:818–827
- Guldner A, Braune A, Carvalho N, Beda A, Zeidler S, Wiedemann B, Wunderlich G, Andreeff M, Uhlig C, Spieth PM, Koch T, Pelosi P, Kotzerke J, de Abreu MG (2014) Higher levels of spontaneous breathing induce lung recruitment and reduce global stress/strain in experimental lung injury. *Anesthesiology* 120:673–682
- Henzler D, Hochhausen N, Bensberg R, Schachtrupp A, Biechele S, Rossaint R, Kuhlens R (2010) Effects of preserved spontaneous breathing activity during mechanical ventilation in experimental intra-abdominal hypertension. *Intensive Care Med* 36:1427–1435
- Hering R, Bolten JC, Kreyer S, Berg A, Wrigge H, Zinserling J, Putensen C (2008) Spontaneous breathing during airway pressure release ventilation in experimental lung injury: effects on hepatic blood flow. *Intensive Care Med* 34:523–527
- Hering R, Viehofer A, Zinserling J, Wrigge H, Kreyer S, Berg A, Minor T, Putensen C (2003) Effects of spontaneous breathing during airway pressure release ventilation on intestinal blood flow in experimental lung injury. *Anesthesiology* 99:1137–1144
- Hering R, Zinserling J, Wrigge H, Varelmann D, Berg A, Kreyer S, Putensen C (2005) Effects of spontaneous breathing during airway pressure release ventilation on respiratory work and muscle blood flow in experimental lung injury. *Chest* 128:2991–2998
- Kill C, Hahn O, Dietz F, Neuhaus C, Schwarz S, Mahling R, Wallot P, Jerrentrup A, Steinfeldt T, Wulf H, Dersch W (2014) Mechanical ventilation during cardiopulmonary resuscitation with intermittent positive-pressure ventilation, bilevel ventilation, or chest compression synchronized ventilation in a pig model. *Crit Care Med* 42:e89–e95
- Kreyer S, Putensen C, Berg A, Soehle M, Muders T, Wrigge H, Zinserling J, Hering R (2010) Effects of spontaneous breathing during airway pressure release ventilation on cerebral and spinal cord perfusion in experimental acute lung injury. *J Neurosurg Anesthesiol* 22:323–329
- Martin LD, Wetzel RC, Bilenki AL (1991) Airway pressure release ventilation in a neonatal lamb model of acute lung injury. *Crit Care Med* 19:373–378
- Matsuzawa Y, Nakazawa K, Yamamura A, Akashi T, Kitagaki K, Eishi Y, Makita K (2010) Airway pressure release ventilation reduces the increase in bronchoalveolar lavage fluid high-mobility group box-1 levels and lung water in experimental acute respiratory distress syndrome induced by lung lavage. *Eur J Anaesthesiol* 27:726–733
- Neumann P, Hedenstierna G (2001) Ventilatory support by continuous positive airway pressure breathing improves gas exchange as compared with partial ventilatory support with airway pressure release ventilation. *Anesth Analg* 92:950–958
- Neumann P, Wrigge H, Zinserling J, Hinz J, Maripuu E, Andersson LG, Putensen C, Hedenstierna G (2005) Spontaneous breathing affects the spatial ventilation and perfusion distribution during mechanical ventilatory support. *Crit Care Med* 33:1090–1095
- Rasanen J, Downs JB, Stock MC (1988) Cardiovascular effects of conventional positive pressure ventilation and airway pressure release ventilation. *Chest* 93:911–915
- Slim AM, Martinho S, Slim J, Davenport E, Castillo-Rojas LM, Shry EA (2011) The effect of airway pressure release ventilation on pulmonary catheter readings: specifically pulmonary capillary wedge pressure in a swine model. *Anesthesiology Res Pract* 2011:371594
- Smith RA, Smith DB (1995) Does airway pressure release ventilation alter lung function after acute lung injury? *Chest* 107:805–808
- Wrigge H, Zinserling J, Neumann P, Defosse J, Magnusson A, Putensen C, Hedenstierna G (2003) Spontaneous breathing improves lung aeration in oleic acid-induced lung injury. *Anesthesiology* 99:376–384

25. Wrigge H, Zinserling J, Neumann P, Muders T, Magnusson A, Putensen C, Hedenstierna G (2005) Spontaneous breathing with airway pressure release ventilation favors ventilation in dependent lung regions and counters cyclic alveolar collapse in oleic-acid-induced lung injury: a randomized controlled computed tomography trial. *Crit Care* 9:R780–R789
26. Xia J, Sun B, He H, Zhang H, Wang C, Zhan Q (2011) Effect of spontaneous breathing on ventilator-induced lung injury in mechanically ventilated healthy rabbits: a randomized, controlled, experimental study. *Crit Care* 15:R244
27. Bratzke E, Downs JB, Smith RA (1998) Intermittent CPAP: a new mode of ventilation during general anesthesia. *Anesthesiology* 89:334–340
28. Cane RD, Peruzzi WT, Shapiro BA (1991) Airway pressure release ventilation in severe acute respiratory failure. *Chest* 100:460–463
29. Chiang AA, Steinfeld A, Gropper C, MacIntyre N (1994) Demand-flow airway pressure release ventilation as a partial ventilatory support mode: comparison with synchronized intermittent mandatory ventilation and pressure support ventilation. *Crit Care Med* 22:1431–1437
30. Dart BW, Maxwell RA, Richart CM, Brooks DK, Ciraulo DL, Barker DE, Burns RP (2005) Preliminary experience with airway pressure release ventilation in a trauma/surgical intensive care unit. *J Trauma* 59:71–76
31. Garner W, Downs JB, Stock MC, Rasanen J (1988) Airway pressure release ventilation (APRV). A human trial. *Chest* 94:779–781
32. Gonzalez M, Arroliga AC, Frutos-Vivar F, Raymonds K, Esteban A, Putensen C, Apezteguia C, Hurtado J, Desmery P, Tomicic V, Elizalde J, Abroug F, Arabi Y, Moreno R, Anzueto A, Ferguson ND (2010) Airway pressure release ventilation versus assist-control ventilation: a comparative propensity score and international cohort study. *Intensive Care Med* 36:817–827
33. Hanna K, Seder CW, Weinberger JB, Sills PA, Hagan M, Janczyk RJ (2011) Airway pressure release ventilation and successful lung donation. *Arch Surg* 146:325–328
34. Hering R, Peters D, Zinserling J, Wrigge H, von Spiegel T, Putensen C (2002) Effects of spontaneous breathing during airway pressure release ventilation on renal perfusion and function in patients with acute lung injury. *Intensive Care Med* 28:1426–1433
35. Kamath SS, Super DM, Mhanna MJ (2010) Effects of airway pressure release ventilation on blood pressure and urine output in children. *Pediatr Pulmonol* 45:48–54
36. Kaplan LJ, Bailey H, Formosa V (2001) Airway pressure release ventilation increases cardiac performance in patients with acute lung injury/adult respiratory distress syndrome. *Crit Care* 5:221–226
37. Liu L, Tanigawa K, Ota K, Tamura T, Yamaga S, Kida Y, Kondo T, Ishida M, Otani T, Sadamori T, Tsumura R, Takeda T, Iwasaki Y, Hirohashi N (2009) Practical use of airway pressure release ventilation for severe ARDS—a preliminary report in comparison with a conventional ventilatory support. *Hiroshima J Med Sci* 58:83–88
38. Maxwell RA, Green JM, Waldrop J, Dart BW, Smith PW, Brooks D, Lewis PL, Barker DE (2010) A randomized prospective trial of airway pressure release ventilation and low tidal volume ventilation in adult trauma patients with acute respiratory failure. *J Trauma* 69:501–510, discussion 511
39. Putensen C, Zech S, Wrigge H, Zinserling J, Stuber F, Von Spiegel T, Mutz N (2001) Long-term effects of spontaneous breathing during ventilatory support in patients with acute lung injury. *Am J Respir Crit Care Med* 164:43–49
40. Rasanen J, Cane RD, Downs JB, Hurst JM, Jousela IT, Kirby RR, Rogove HJ, Stock MC (1991) Airway pressure release ventilation during acute lung injury: a prospective multicenter trial. *Crit Care Med* 19:1234–1241
41. Schultz TR, Costarino AJA, Durning SM, Napoli LA, Schears G, Godinez RI, Priestley M, Dominguez T, Lin R, Helfaer M (2001) Airway pressure release ventilation in pediatrics. *Pediatr Crit Care Med* 2:243–246
42. Sydow M, Burchardi H, Ephraim E, Zielmann S, Crozier TA (1994) Long-term effects of two different ventilatory modes on oxygenation in acute lung injury. Comparison of airway pressure release ventilation and volume-controlled inverse ratio ventilation. *Am J Respir Crit Care Med* 149:1550–1556
43. Varpula T, Jousela I, Niemi R, Takkunen O, Pettila V (2003) Combined effects of prone positioning and airway pressure release ventilation on gas exchange in patients with acute lung injury. *Acta Anaesthesiol Scand* 47:516–524
44. Varpula T, Valta P, Niemi R, Takkunen O, Hynynen M, Pettila VV (2004) Airway pressure release ventilation as a primary ventilatory mode in acute respiratory distress syndrome. *Acta Anaesthesiol Scand* 48:722–731
45. Wrigge H, Zinserling J, Hering R, Schwalfenberg N, Stuber F, von Spiegel T, Schroeder S, Hedenstierna G, Putensen C (2001) Cardiorespiratory effects of automatic tube compensation during airway pressure release ventilation in patients with acute lung injury. *Anesthesiology* 95:382–389
46. Maung AA, Luckianow G, Kaplan LJ (2012) Lessons learned from airway pressure release ventilation. *J Trauma Acute Care Surg* 72:624–628
47. Maung AA, Schuster KM, Kaplan LJ, Ditillo MF, Piper GL, Maerz LL, Lui FY, Johnson DC, Davis KA (2012) Compared to conventional ventilation, airway pressure release ventilation may increase ventilator days in trauma patients. *J Trauma Acute Care Surg* 73:507–510
48. Testerman GM, Breitman I, Hensley S (2013) Airway pressure release ventilation in morbidly obese surgical patients with acute lung injury and acute respiratory distress syndrome. *Am Surg* 79:242–246
49. Albert S, Kubiak BD, Vieau CJ, Roy SK, DiRocco J, Gatto LA, Young JL, Tripathi S, Trikha G, Lopez C, Nieman GF (2011) Comparison of “open lung” modes with low tidal volumes in a porcine lung injury model. *J Surg Res* 166:e71–e81
50. Andrews PL, Shiber JR, Jaruga-Killeen E, Roy S, Sadowitz B, O’Toole RV, Gatto LA, Nieman GF, Scalet T, Habashi NM (2013) Early application of airway pressure release ventilation may reduce mortality in high-risk trauma patients: a systematic review of observational trauma ARDS literature. *J Trauma Acute Care Surg* 75:635–641
51. Arrindell EL Jr, Krishnan R, van der Merwe M, Caminita F, Howard SC, Zhang J, Buddington RK (2015) Lung volume recruitment in a preterm pig model of lung immaturity. *Am J Physiol Lung Cell Mol Physiol* 309: L1088–L1092
52. Davies SW, Leonard KL, Falls RK Jr, Mageau RP, Efrid JT, Hollowell JP, Trainor WE 2nd, Kanaan HA, Hickner RC, Sawyer RG, Poulin NR, Waibel BH, Toschlog EA (2015) Lung protective ventilation (ARDSNet) versus airway

- pressure release ventilation: ventilatory management in a combined model of acute lung and brain injury. *J Trauma Acute Care Surg* 78:240–249, discussion 249–251
53. Kawaguchi A, Guerra GG, Duff JP, Ueta I, Fukushima R (2014) Hemodynamic changes in child acute respiratory distress syndrome with airway pressure release ventilation: a case series. *Clin Respir J*. doi:10.1111/crj.12155. [Epub ahead of print]
  54. Kollisch-Singule M, Emr B, Smith B, Roy S, Jain S, Satalin J, Snyder K, Andrews P, Habashi N, Bates J, Marx W, Nieman G, Gatto LA (2014) Mechanical breath profile of airway pressure release ventilation: the effect on alveolar recruitment and microstrain in acute lung injury. *JAMA Surg* 149:1138–1145
  55. Kollisch-Singule M, Emr B, Smith B, Ruiz C, Roy S, Meng Q, Jain S, Satalin J, Snyder K, Ghosh A, Marx W, Andrews P, Habashi N, Nieman G, Gatto LA (2014) Airway pressure release ventilation reduces conducting airway micro-strain in lung injury. *J Am Coll Surg* 219:9
  56. Kollisch-Singule M, Jain S, Andrews P, Smith BJ, Hamlington-Smith KL, Roy S, DiStefano D, Nuss E, Satalin J, Meng Q, Marx W, Bates JH, Gatto LA, Nieman GF, Habashi NM (2016) Effect of airway pressure release ventilation on dynamic alveolar heterogeneity. *JAMA Surg* 151(1):64–72.
  57. Roy S, Sadowitz B, Andrews P, Gatto LA, Marx W, Ge L, Wang G, Lin X, Dean DA, Kuhn M, Ghosh A, Satalin J, Snyder K, Vodovotz Y, Nieman G, Habashi N (2012) Early stabilizing alveolar ventilation prevents acute respiratory distress syndrome: a novel timing-based ventilatory intervention to avert lung injury. *J Trauma Acute Care Surg* 73:391–400
  58. Walsh MA, Merat M, La Rotta G, Joshi P, Joshi V, Tran T, Jarvis S, Caldaroni CA, Van Arsdell GS, Redington AN, Kavanagh BP (2011) Airway pressure release ventilation improves pulmonary blood flow in infants after cardiac surgery. *Crit Care Med* 39:2599–2604
  59. Yehya N, Topjian AA, Lin R, Berg RA, Thomas NJ, Friess SH (2014) High frequency oscillation and airway pressure release ventilation in pediatric respiratory failure. *Pediatr Pulmonol* 49:707–715
  60. Yoshida T, Rinka H, Kaji A, Yoshimoto A, Arimoto H, Miyaichi T, Kan M (2009) The impact of spontaneous ventilation on distribution of lung aeration in patients with acute respiratory distress syndrome: airway pressure release ventilation versus pressure support ventilation. *Anesth Analg* 109:1892–1900
  61. Roy SK, Emr B, Sadowitz B, Gatto LA, Ghosh A, Satalin JM, Snyder KP, Ge L, Wang G, Marx W, Dean D, Andrews P, Singh A, Scalea T, Habashi N, Nieman GF (2013) Preemptive application of airway pressure release ventilation prevents development of acute respiratory distress syndrome in a rat traumatic hemorrhagic shock model. *Shock* 40:210–216
  62. Emr B, Gatto LA, Roy S, Satalin J, Ghosh A, Snyder K, Andrews P, Habashi N, Marx W, Ge L, Wang G, Dean DA, Vodovotz Y, Nieman G (2013) Airway pressure release ventilation prevents ventilator-induced lung injury in normal lungs. *JAMA Surg* 148:1005–1012
  63. (2000) Ventilation with lower tidal volumes as compared with traditional tidal volumes for acute lung injury and the acute respiratory distress syndrome. The acute respiratory distress syndrome network. *N Engl J Med* 342:1301–1308
  64. Protti A, Andreis DT, Monti M, Santini A, Sparacino CC, Langer T, Votta E, Gatti S, Lombardi L, Leopardi O, Masson S, Cressoni M, Gattinoni L (2013) Lung stress and strain during mechanical ventilation: any difference between statics and dynamics? *Crit Care Med* 41:1046–1055
  65. Gattinoni L, Protti A, Caironi P, Carlesso E (2010) Ventilator-induced lung injury: the anatomical and physiological framework. *Crit Care Med* 38:S539–S548
  66. Retamal J, Bergamini BC, Carvalho AR, Bozza FA, Borzone G, Borges JB, Larsson A, Hedenstierna G, Bugedo G, Bruhn A (2014) Non-lobar atelectasis generates inflammation and structural alveolar injury in the surrounding healthy tissue during mechanical ventilation. *Crit Care* 18:505
  67. Caironi P, Cressoni M, Chiumello D, Ranieri M, Quintel M, Russo SG, Cornejo R, Bugedo G, Carlesso E, Russo R, Caspani L, Gattinoni L (2010) Lung opening and closing during ventilation of acute respiratory distress syndrome. *Am J Respir Crit Care Med* 181:578–586
  68. Rose L, Hawkins M (2008) Airway pressure release ventilation and biphasic positive airway pressure: a systematic review of definitional criteria. *Intensive Care Med* 34:1766–1773
  69. Facchin F, Fan E (2015) Airway pressure release ventilation and high-frequency oscillatory ventilation: potential strategies to treat severe hypoxemia and prevent ventilator-induced lung injury. *Respir Care* 60:1509–1521

**Submit your manuscript to a SpringerOpen<sup>®</sup> journal and benefit from:**

- Convenient online submission
- Rigorous peer review
- Immediate publication on acceptance
- Open access: articles freely available online
- High visibility within the field
- Retaining the copyright to your article

---

Submit your next manuscript at ► [springeropen.com](http://springeropen.com)

---