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# Can linezolid be validly measured in endotracheal aspiration in critically ill patients? A proof-of-concept trial



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# Abstract

**Background** Therapeutic drug monitoring (TDM) of anti-infectives such as linezolid is routinely performed in blood of intensive care unit (ICU) patients to optimize target attainment. However, the concentration at the site of infection is considered more important for a successful therapy. Until now, bronchoalveolar lavage (BAL) is the gold standard to measure intrapulmonary concentrations of anti-infective agents. However, it is an invasive method and unsuitable for regular TDM. The aim of this proof-of-concept study was to investigate whether it is possible to reliably determine the intrapulmonary concentration of linezolid from endotracheal aspiration (ENTA).

**Methods** Intubated ICU patients receiving 600 mg intravenous linezolid twice daily were examined in steady state. First, preliminary experiments were performed in six patients to investigate which patients are suitable for linezolid measurement in ENTA. In a second step, trough and peak linezolid concentrations of plasma and ENTA were determined in nine suitable patients.

**Results** Linezolid can validly be detected in ENTA with viscous texture and > 0.5 mL volume. The mean (SD) linezolid trough concentration was 2.02 (1.27) mg/L in plasma and 1.60 (1.36) mg/L in ENTA, resulting in a median lung penetration rate of 104%. The mean (SD) peak concentration in plasma and ENTA was 10.77 (5.93) and 4.74 (2.66) mg/L.

**Conclusions** Linezolid can validly be determined in ENTA with an adequate texture and volume. The penetration rate is comparable to already published BAL concentrations. This method might offer a simple and non-invasive method for TDM at the site of infection "lung". Due to promising results of the feasibility study, comparison of ENTA and BAL in the same patient should be investigated in a further trial.

**Keywords** Linezolid, Critically ill patient, Endotracheal aspiration (ENTA), Therapeutic drug monitoring, Bronchoalveolar lavage (BAL)

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# Background

Infections are a major challenge in intensive care units. 54% of patients in intensive care units have a confirmed infection and 70% receive at least one antibiotic [1]. Furthermore, the 30-day mortality rate for septic shock in Germany is as high as 30% despite different approaches to optimize therapy [2]. Anti-infective therapy has, therefore, a crucial role in the treatment of patients with severe infections. However, the dosing of anti-infectives is more difficult in intensive care unit (ICU) patients, as they



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have altered pharmacokinetics due to profoundly altered pathophysiological processes [3, 4]. Routine therapeutic drug monitoring (TDM) of especially antibiotics in ICU patients is recommended to reduce the risk of under- or overdosing, to maximize the efficacy, and to minimize toxicity [5–7], hopefully leading to a higher rate of target attainment.

Linezolid is regularly used in ICU patients with pneumonia or skin and soft tissue infections caused by gram-positive bacteria, such as methicillin-resistant Staphylococcus aureus (MRSA) or vancomycin-resistant enterococci (VRE) [8]. Large inter- and intra-individual differences in blood concentrations of linezolid were observed, leading to a complex dosing strategy [9]. Furthermore, the concentration in the blood does not automatically indicate the concentration at the site of infection, which is hypothesized to be more relevant for a successful therapy [10]. It has been shown that patients with acute respiratory distress syndrome or pneumonia have significantly lower linezolid blood concentrations than patients with non-pulmonary infections [11, 12]. Moreover, linezolid has been described to have a high penetration rate into the lung tissue [13]. It is not vet understood which linezolid concentration should be aimed for in the blood to achieve adequate concentrations in the lungs.

The gold standard for determining the intrapulmonary concentration of anti-infectives such as linezolid is the measurement in the epithelial lining fluid (ELF) obtained during bronchoscopy with bronchoalveolar lavage (BAL) [14]. However, bronchoscopy is an invasive procedure and not suitable for routine TDM as it, i.e., leads to a loss of positive end-expiratory pressure (PEEP) [15]. In addition, the secretion in the BAL is diluted with saline, so the TDM methods must be developed with a considerably lower calibration range and the actual concentration is calculated using the urea method, which is also prone to error [16].

The aim of this proof-of-concept study was to evaluate whether a reliable quantitative linezolid determination from endotracheal aspiration (ENTA) is possible. If possible, this would provide a simple and non-invasive method for TDM of linezolid at the site of infection "lung", which would be an important step towards personalized dosing of anti-infectives.

## Methods

# Study design

This prospective, monocentric, proof-of-concept study was conducted at the LMU Hospital in Munich in two anesthesiologic ICUs. The local review board (project number 22-0490) gave ethics approval. The study was registered in the German Register for Clinical Studies (DRKS00030870).

Intubated, adult ICU patients receiving 600 mg intravenously linezolid twice daily, were included. Informed consent for the study was obtained from the patients or their legal representative. First, preliminary experiments were performed to define and develop the conditions and methodology for a TDM of linezolid from ENTA. Thereafter, the trough and peak concentration of linezolid in ENTA and plasma were determined in patients considered suitable for ENTA sampling and linezolid TDM. All demographic and clinical data were obtained from the patients' medical records.

## Sampling, processing, and measurement

The steady-state trough and peak levels were used to compare the concentrations in ENTA and plasma. The ENTA was obtained in a standardized manner. Whenever possible, the patient was not suctioned for 1 h before sampling and no inhalatives were administered 1 h before sampling. A closed suction system (14Ch, suction depth 54 cm) with the same negative pressure (-20 mmHg)was always used. Linezolid was quantified from plasma that was obtained from arterial blood and ENTA. The linezolid concentration in plasma was determined using an established routine isotope-dilution liquid chromatography-tandem mass spectrometry (ID LC-MS/MS) method with a lower limit of quantification of 0.125 mg/L [17]. To qualify ENTA for linezolid measurement with the very same ID LC-MS/MS method, the viscous secretion had first to be liquefied. The aspirate was diluted 1:2 (v/v) with Proteinase-K solution (1 mg/mL Proteinase-K in 50 mM Tris-HCL, 200 mM CaCl2, total pH 7.5), and after short vortexing, the mixture was incubated in a water bath at 37 °C for 20 min. Preliminary experiments confirmed stability of linezolid in ENTA at 37 °C for 90 min (deviation  $< \pm 15\%$ ). The plasma and liquified ENTA were stably stored up to 1 month at -80 °C until ID LC-MS/MS analysis.

# Statistical analysis

Statistical analysis was performed using IBM SPSS Statistics (version 29.0.0.0). Normal distribution was tested using the Shapiro–Wilk test. Values were expressed as mean and standard deviation (SD) or as median and interquartile range (IQR). Pearson correlation was used to correlate trough levels of ENTA and plasma.

## Results

Preliminary experiments were performed in six patients to define and develop the conditions and methodology for a TDM from ENTA. In five out of six patients, the ENTA volume was  $\leq 0.5$  mL and/ or had a liquid texture,

 Table 1
 ENTA volume, texture, trough concentration, and site of infection of preliminary experiments

Patient	Volume (mL)	Texture	Trough concentration ENTA (mg/L)	Site of Infection
1	0.4	Viscous	< 0.125	Abdomen
2	0.3	Liquid	1.26	Abdomen
3	0.2	Liquid	< 0.125	Extremity
4	0.8	Viscous	0.92	Abdomen + lung
5	0.5	Liquid	< 0.125	Abdomen
6	1.0	Liquid	< 0.125	Extremity

 Table 2
 Patient characteristics

Patient characteristics	n (%)	Mean (SD)	Median (IQR)
Number of patients	9 (100)		
Male	8 (88.9)		
Age (years)		59 (17)	
Body size (cm)		183 (10)	
Body weight (kg)			86 (25.5)
SAPS II at start linezolid therapy		63 (15)	
APACHE II on admission to ICU		26 (8)	
Length of stay in ICU (days)		47 (18)	
Indication linezolid: E. faecalis	5 (55.6)		
Indication linezolid: calculated	4 (44.4)		
Proven pulmonary infection	7 (77.8)		
28-day mortality	2 (22.2)		

SAPS simplified acute physiology score, APACHE acute physiology and chronic health evaluation

suggesting the hypothesis of condensate accumulation. Linezolid could not or not validly be detected in these samples. Table 1 displays the ENTA volume and texture as well as the measured trough concentration in the ENTA and the site of infection.

Thereafter, only patients with a viscous aspirate > 0.5 mL were included and a total of nine patients was observed for the proof-of-concept trial. Seven patients had a proven pulmonary infection confirmed by either image morphology or microbial detection. Patients 2, 4, 5, 6, 7, 8, and 9 were treated with inhalatives (71% Amphotericin B, 71% Ipratropium bromid + Salbutamol, 57% Sodium chloride 3%, 14% Colistin). The reason for admission to the ICU was in descending order: respiratory failure/ARDS (n=4), sepsis/septic shock (n=3), polytrauma (n=1), liver transplantation (n=1). Table 2 shows further patient characteristics.

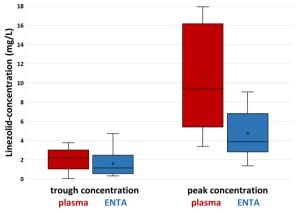
Linezolid trough and peak concentrations were measured in plasma and ENTA that are displayed in Table 3. In patients 1, 3, and 6, sufficient viscous secretion was used to determine trough concentrations of plasma and ENTA at two timepoints.

The mean (SD) linezolid trough levels were 2.02 (1.27) mg/L in plasma and 1.60 (1.36) mg/L in ENTA. The median ENTA/plasma ratio was 104.2%. Pearson correlation showed a significant and moderate correlation between plasma and ENTA trough levels (r=0.52, p < 0.05). The mean (SD) linezolid peak concentrations were 10.77 (5.93) mg/L in plasma and 4.74 (2.66) mg/L in ENTA with a median penetration rate of 47.7%. Figure 1 illustrates the linezolid trough and peak concentrations in plasma and ENTA.

ENTA volume	Trough conc. plasma (mg/L)	Trough conc. ENTA (mg/L)	Ratio ENTA/plasma (%)	Peak conc. plasma (mg/L)	Peak conc. ENTA (mg/L)
0.8/0.7	3.78 / 3.22	0.92 / 0.34	24.3 / 10.6	15.59	2.82
0.9	0.05	0.48	960.0	3.39	2.80
3.2/1.3	2.94 / 3.15	2.42 / 4.72	82.3 / 149.8	7.45	4.52
1.5	0.31	0.32	103.2	5.41	1.36
0.6	2.98	3.16	106.0	16.17	6.82
1.3/0.6	1.21 / 1.88	1.38 / 1.08	114.1 / 57.4	9.37	3.24
2.3	0.52	0.58	111.5	4.42	3.88
2.0	2.49	2.62	105.2	17.92	9.04
0.9	1.69	1.22	72.2	17.19	8.20
	2.02 (1.27)	1.60 (1.36)		10.77 (5.93)	4.74 (2.66)
			104.2 (43.6)		
	0.8/0.7 0.9 3.2/1.3 1.5 0.6 1.3/0.6 2.3 2.0	(mg/L) 0.8/0.7 3.78 / 3.22 0.9 0.05 3.2/1.3 2.94 / 3.15 1.5 0.31 0.6 2.98 1.3/0.6 1.21 / 1.88 2.3 0.52 2.0 2.49 0.9 1.69	(mg/L)         (mg/L)           0.8/0.7         3.78 / 3.22         0.92 / 0.34           0.9         0.05         0.48           3.2/1.3         2.94 / 3.15         2.42 / 4.72           1.5         0.31         0.32           0.6         2.98         3.16           1.3/0.6         1.21 / 1.88         1.38 / 1.08           2.3         0.52         0.58           2.0         2.49         2.62           0.9         1.69         1.22	(mg/L)         (mg/L)         ENTA/plasma (%)           0.8/0.7         3.78 / 3.22         0.92 / 0.34         24.3 / 10.6           0.9         0.05         0.48         960.0           3.2/1.3         2.94 / 3.15         2.42 / 4.72         82.3 / 149.8           1.5         0.31         0.32         103.2           0.6         2.98         3.16         106.0           1.3/0.6         1.21 / 1.88         1.38 / 1.08         114.1 / 57.4           2.3         0.52         0.58         111.5           2.0         2.49         2.62         105.2           0.9         1.69         1.22         72.2           2.02 (1.27)         1.60 (1.36)         1.21	(mg/L)         (mg/L)         ENTA/plasma (%)         plasma (mg/L)           0.8/0.7         3.78 / 3.22         0.92 / 0.34         24.3 / 10.6         15.59           0.9         0.05         0.48         960.0         3.39           3.2/1.3         2.94 / 3.15         2.42 / 4.72         82.3 / 149.8         7.45           1.5         0.31         0.32         103.2         5.41           0.6         2.98         3.16         106.0         16.17           1.3/0.6         1.21 / 1.88         1.38 / 1.08         114.1 / 57.4         9.37           2.3         0.52         0.58         111.5         4.42           2.0         2.49         2.62         105.2         17.92           0.9         1.69         1.22         72.2         10.77 (5.93)

**Table 3** Linezolid trough and peak concentrations in plasma and ENTA

\*No proven pneumonia



**Fig. 1** Linezolid trough and peak concentration in plasma and ENTA. The boxes of the boxplots represent the interquartile range (IQR), the line the median, the cross the mean and the whiskers the minimum and maximum value. The red boxplots represent the trough and peak concentrations in plasma and the blue boxplots represent the trough concentrations in ENTA

If sufficient secretions could be aspirated, further linezolid measurements were carried out in the ENTA and in the blood. Figure 2 shows the linezolid concentration of the 9 patients in the ENTA (blue) and plasma (red) over the 12-h dosing interval.

# Discussion

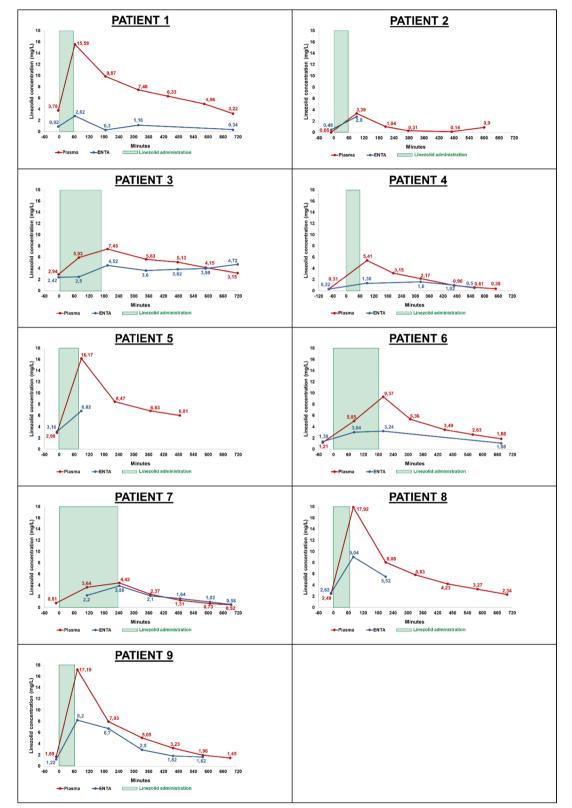
The concentration of antibiotics in the blood does not automatically indicate the concentration at the site of infection. Linezolid is often used in critically ill patients with pneumonia due to a postulated high penetration rate and is associated with a better outcome than the treatment with vancomycin [13, 18]. The measurement of linezolid in patients' lung, if the suffer from pneumonia, seems, therefore, promising [14, 19]. The high rate of *E. faecalis* in our collective might be caused by the high number of immunosuppressed patients after severe infections with prolonged intensive care stay or organ transplantation. In other hospitals, the detection of, e.g., pneumococci, is to be expected.

Measuring linezolid concentrations in endotracheal aspirate is a potential new, simpler, and non-invasive method compared to BAL. It would be less time-consuming and does not require additional equipment, such as a bronchoscope. If the suctioning is performed with a closed system, the patient is not exposed to any loss of PEEP or risk of infection [15]. However, the suction depth, the thickness of the catheter and a potential biofilm formation in the endotracheal tubes could have an influence on the quality of the sample and the included linezolid concentration. In contrast, a standardized BAL also has different limitations: the dwell time plays a central role in the percentage of the drug that is transferred into the aspirate [20]. Furthermore, the high dilution with NaCl leads to very low drug concentrations with the necessity of reevaluating the calibration range. The transfer factor is typically estimated using the urea method, which, as already mentioned, is only an estimate [21, 22].

The preliminary measurements taught us that validly measurement of linezolid in the ENTA is possible in samples with sufficient volume (>0.5 mL) and viscous texture. If there is too little liquid secretion, it is possible that only the condensation water from the ventilation tube is drawn off, and therefore, no valid measurement is possible. Patients with pneumonia mostly have viscous pulmonary secretions and the linezolid concentration in the lung is especially of interest in those patients. Linezolid trough concentrations in ENTA were shown to be comparable to plasma concentrations in appropriate patients, as confirmed by a moderate correlation coefficient of 0.52. In fact, 58% of the trough levels were even higher in ENTA than in plasma, which has already been observed in the BAL, demonstrating the good penetration of linezolid into the lung and indicating that our method might be valid for non-invasive measurement of linezolid in the critically ill [19, 23].

Although also peak concentrations were measured in plasma and ENTA and further concentrations in the dosing interval, no reliable interpretation can be made due to the lack of knowledge about the penetration velocity in patients with intermitted dosing. To improve the interpretation of the data, a pharmacokinetic model would be beneficial in the future. Significant differences were observed in the lung penetration rate of linezolid (e.g., patient 1 with a very low penetration rate). No specific cause can be named for this, although large inter-individual differences in the linezolid blood concentration are already known and the reasons for this are at least partially transferable to the lungs [9, 12]. This again demonstrates the need for TDM of linezolid not only in the blood but also at the site of infection to identify patients like patient 1 with low penetration rates and to enable the target range to be achieved in these patients as well [3, 21, 24].

Only four published studies were identified that investigated the intrapulmonary trough concentration of intravenously administered linezolid in critically ill patients using the gold standard "BAL". This already shows that the current data situation is very limited. Boselli et al. studied the intrapulmonary concentration of linezolid in 16 patients with ventilator-associated pneumonia. The mean linezolid penetration into ELF was 104% [25]. In another study, Boselli et al. investigated the alveolar diffusion of linezolid during continuous linezolid administration in



**Fig. 2** Linezolid concentration in ENTA and plasma over the 12-h dosing interval. The red line represents the concentration of linezolid in the plasma and the blue line the concentration in the ENTA. The green box shows the administration time of linezolid. Only ENTA values with a volume > 0.5 mL and a viscous texture are shown

12 patients with a median (IQR) linezolid penetration rate of 97% [19]. De Pascale et al. received a median penetration rate of 80% in seven critically ill obese patients [14]. Finally, Wu et al. studied 23 patients with sepsis and was able to show a penetration rate of 112% [10]. The median penetration rate into the ELF in these four studies is 101%. Based on the available data, all studies demonstrate a low variability of the penetration rate.

We observed a median ENTA/plasma ratio of the trough concentrations of 104%. The penetration rate measured in our study appears to be comparable to the median penetration rate measured in the ELF in the above-mentioned studies. It should be noted that a direct comparison is not possible as different patients were used. This allows the hypothesis to be derived that the measurement of linezolid in the ENTA might be a valid method for the future. To test this hypothesis, the measurement of linezolid in ENTA and BAL in the same patient at the same time is pending. The primary goal of examining whether linezolid can be measured in endotracheal aspiration in patients with viscous secretions was thus achieved.

This proof-of-concept trial has several limitations. Only nine patients were included and, therefore, less patients as in the published studies where linezolid was measured in the ELF. However, the goal of the proof of concept study was to verify if and when a measurement of linezolid in the ENTA is possible at all. This goal was achieved by including nine selected and valuable patients. Since the speed of linezolid penetration into the lungs is not yet known, it is unclear whether the linezolid peak levels in the blood corresponds to the peak level in the lungs when measured at the same time. A pharmacokinetic model could, therefore, be helpful. The penetration of linezolid into pneumonic areas may be limited, as they are less perfused under the assumption of the Euler-Liljestrand reflex. A BAL directly from this area might contribute to an even more precise determination of the concentration at the site of infection. Finally, linezolid in the lung was only determined in ENTA and not additionally from the BAL in the same patient. A direct comparison of the concentrations is not possible yet. The next step should be to directly compare the concentration in the ENTA with the concentration in the BAL in the same patient at the same time. Thereafter, ENTA can perhaps be used as a non-invasive standard method in the future.

# Conclusion

Linezolid can be reliably quantified in ENTA with adequate texture and volume. This might offer a new and simple way to determine the concentration of linezolid for TDM at the site of infection "lung", allowing even more personalized and targeted therapy for suitable and selected patients. In the future, validation of the method with the gold standard "BAL" for measuring the intrapulmonary concentration is recommended.

#### Abbreviations

BAL	Bronchoalveolar lavage				
ELF	Epithelial lining fluid				
ENTA	Endotracheal aspirates				
ICU	Intensive care unit				
ID LC–MS/MS	Isotope dilution liquid chromatography-tandem mass				
	spectrometry				
PEEP	Positive end-expiratory pressure				
TDM	Therapeutic drug monitoring				

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#### Author contributions

CS, UL, MP, and KH designed the study. CG and JP screened the patients for study inclusion. DR and ClB recruited the patients. MP and KH developed the method for liquefying the ENTA. MP, KH, LFH and MV assayed the samples. DR wrote the manuscript. All authors meet key authorship requirements and agree to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. All authors read and approved the final manuscript.

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#### Availability of data and materials

Linezolid was determined at more measurement times than the trough level, but no further conclusions can be drawn from the descriptive analysis.

## Declarations

#### Ethics approval and consent to participate

Ethical approval was obtained from the ethical review committee of the Ludwig-Maximilians-Universität (Registration Number 22–0490).

#### **Consent for publication**

Not applicable.

#### **Competing interests**

The authors declare that they have no competing interests.

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