

REVIEW

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Pathophysiology and clinical consequences of arterial blood gases and pH after cardiac arrest



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Abstract

Post cardiac arrest syndrome is associated with high morbidity and mortality, which is related not only to a poor neurological outcome but also to respiratory and cardiovascular dysfunctions. The control of gas exchange, and in particular oxygenation and carbon dioxide levels, is fundamental in mechanically ventilated patients after resuscitation, as arterial blood gases derangement might have important effects on the cerebral blood flow and systemic physiology.

In particular, the pathophysiological role of carbon dioxide (CO₂) levels is strongly underestimated, as its alterations quickly affect also the changes of intracellular pH, and consequently influence metabolic energy and oxygen demand. Hypo/hypercapnia, as well as mechanical ventilation during and after resuscitation, can affect CO₂ levels and trigger a dangerous pathophysiological vicious circle related to the relationship between pH, cellular demand, and catecholamine levels. The developing hypocapnia can nullify the beneficial effects of the hypothermia. The aim of this review was to describe the pathophysiology and clinical consequences of arterial blood gases and pH after cardiac arrest.

According to our findings, the optimal ventilator strategies in post cardiac arrest patients are not fully understood, and oxygen and carbon dioxide targets should take in consideration a complex pattern of pathophysiological factors. Further studies are warranted to define the optimal settings of mechanical ventilation in patients after cardiac arrest.

Keywords: Cardiac arrest, Ventilator targets, Intracellular acidosis, Catecholamine, Gas exchanges

Background

The aim of cardiopulmonary resuscitation is to promptly restore spontaneous circulation in order to avoid hypoxic ischemic brain injury. In this context, ventilation has the goal to maintain an appropriate level of arterial oxygen (PaO_2), while reducing arterial carbon dioxide levels (PaCO_2), avoiding respiratory acidosis. However, the pathophysiology of cardiac arrest and its systemic effects, as well as the relationship between arterial blood gases, hemodynamic management, and ventilation during and after resuscitation is complex. Evidence regarding the optimal oxygen or carbon dioxide (CO_2) targets is limited, and the relationship between PaO_2 and PaCO_2 targets in the context of catecholamine administration and pH changes is not fully understood. Also, the pathophysiological effects of CO_2 are underestimated in the literature; variations of carbon dioxide levels can quickly change the extracellular-intracellular concentration of H^+ , and modifications in pH and acidosis have extraordinary importance on systemic homeostasis [1].

The aim of this review is to discuss the pathophysiology of arterial blood gases and pH in mechanically ventilated patients after cardiac arrest and their clinical consequences. We will not focus on ventilator settings but on the pH, PaO_2 , and CO_2 physiological targets to achieve in this cohort of patients.

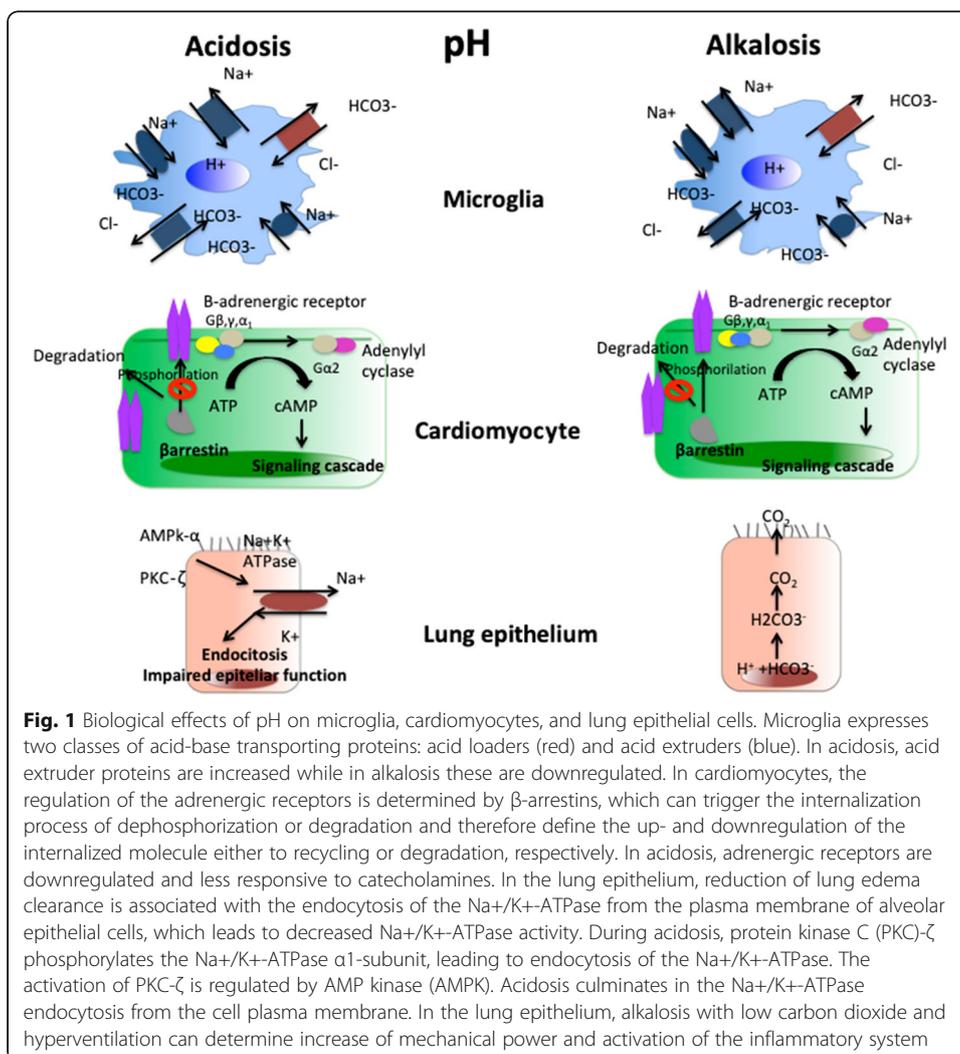


Fig. 1 Biological effects of pH on microglia, cardiomyocytes, and lung epithelial cells. Microglia expresses two classes of acid-base transporting proteins: acid loaders (red) and acid extruders (blue). In acidosis, acid extruder proteins are increased while in alkalosis these are downregulated. In cardiomyocytes, the regulation of the adrenergic receptors is determined by β -arrestins, which can trigger the internalization process of dephosphorization or degradation and therefore define the up- and downregulation of the internalized molecule either to recycling or degradation, respectively. In acidosis, adrenergic receptors are downregulated and less responsive to catecholamines. In the lung epithelium, reduction of lung edema clearance is associated with the endocytosis of the Na^+/K^+ -ATPase from the plasma membrane of alveolar epithelial cells, which leads to decreased Na^+/K^+ -ATPase activity. During acidosis, protein kinase C (PKC)- ζ phosphorylates the Na^+/K^+ -ATPase $\alpha 1$ -subunit, leading to endocytosis of the Na^+/K^+ -ATPase. The activation of PKC- ζ is regulated by AMP kinase (AMPK). Acidosis culminates in the Na^+/K^+ -ATPase endocytosis from the cell plasma membrane. In the lung epithelium, alkalosis with low carbon dioxide and hyperventilation can determine increase of mechanical power and activation of the inflammatory system

Pathophysiological basis

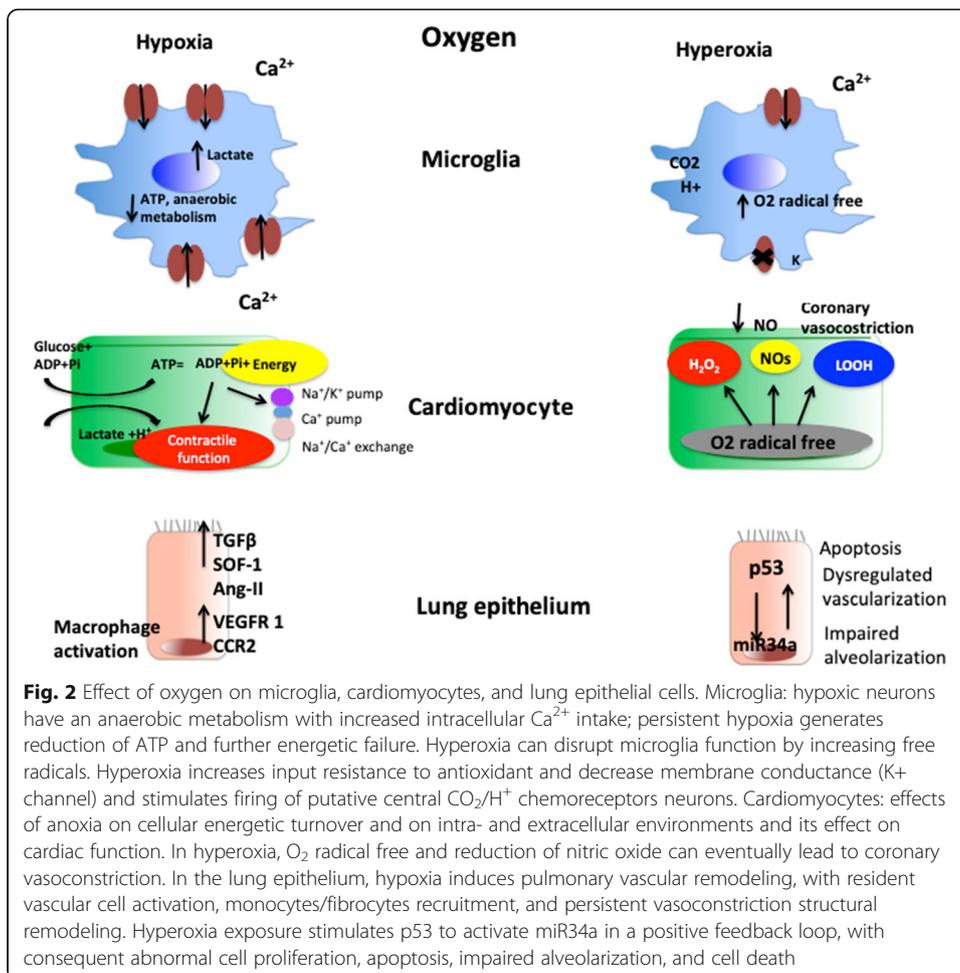
Acidosis and lactate

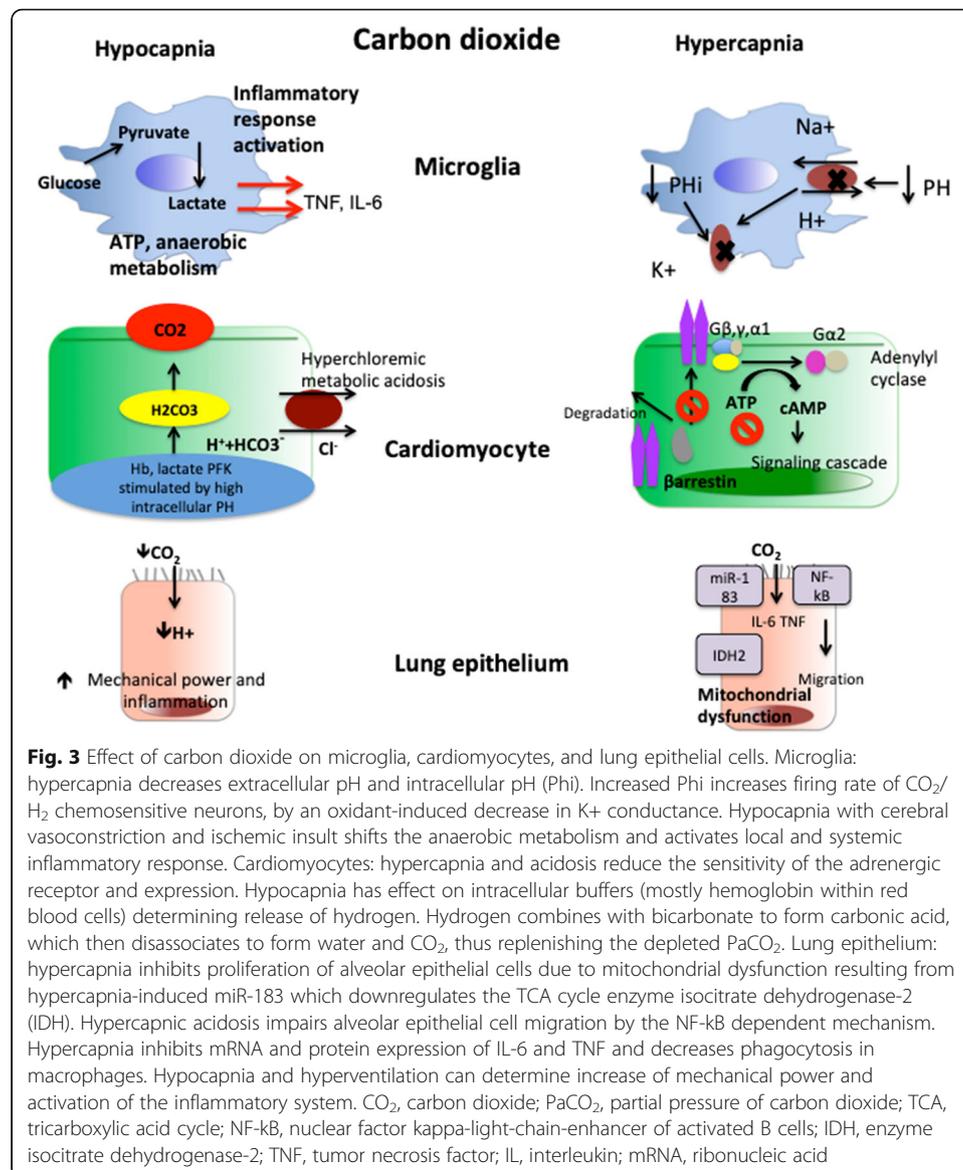
Acute circulatory shock and cardiac arrest predispose to inadequate oxygen delivery, resulting in cellular hypoxia and acute brain damage [2] (Figs. 1, 2, 3).

Patients after cardiac arrest may often present with a combination of metabolic and respiratory disturbances, where lactic acidosis is the main cause of metabolic acidosis (defined as $\text{pH} \leq 7.35$ and lactatemia $> 2.0 \text{ mmol l}^{-1}$ with a $\text{PaCO}_2 \leq 42 \text{ mmHg}$) [3, 4].

Hypoperfusion and acidotic disturbances during return of spontaneous circulation (ROSC) result in a misbalance between lactate production and removal; the dissociation of lactic acid produces hydrogen ions which are used for the oxidative phosphorylation and adenosine Tri-Phosphate (ATP) production [3, 5]. In a canine model, Sanders et al. [6] showed that metabolic acidosis is predominant in the late phase of cardiac arrest, while in the first short period after circulatory arrest, respiratory alkalosis is more frequent, probably related to artificial hyperventilation.

However, acid-base disturbances imply a larger spectrum of clinical consequences. Acute lactic acidosis depresses left ventricular contractility, triggers the release of endogenous catecholamine, and decreases the responsiveness of the left ventricle to the





actions of catecholamine [7]. Therefore, late detection and treatment of acidosis in the context of impaired cardiovascular function and responsiveness to vasopressors in post cardiac arrest status may lead to worse outcome [8].

The pathophysiological effects of acidosis on cardiomyocytes are well described [2]; extracellular and intracellular acidosis reduces the number of adrenoreceptors expressed on the cell membrane, in particular beta receptors, and contributes to reduced contractile responsiveness to catecholamines [9]. Decreased pH also influences the synthesis and release of tumor necrosis factor (TNF- α) and nitric oxide (NO) in macrophages or macrophage-like cell lines and can “per se” affect hemodynamic stability: moderate acidosis (pH = 7–7.20) might elevate nitric oxide (NO) production and levels, but severe acidosis (pH < 7.0) determines a decrease in NO release and NO synthesis [10–13].

Oxygen

After successful resuscitation and ROSC, achieving appropriate levels of oxygen and carbon dioxide in arterial blood is necessary to avoid further hypoxia and secondary damage [14] (Figs. 1, 2, 3). A large cohort study performed in 82 intensive care units (ICUs) in hospitals participating to the Dutch National Intensive Care Evaluation (NICE) [15] showed that both PaCO₂ and PaO₂ levels in post cardiac arrest patients are predictors for hospital mortality.

Re-oxygenation after the no flow phase is a major stimulant for myocardial catecholamine release [16]. Decreased arterial O₂ levels are associated with low pH and almost immediate sodium-potassium (Na⁺/K⁺) ATPase dysfunction, with consequent ion imbalance. Thus, hypoxemia induces mitochondrial damage, which determinates systemic inflammation with energy failure and cell apoptosis [17]. On the other hand, even hyperoxia can be detrimental and has shown to be associated with higher mortality (EMShockNet investigators) [18].

Hyperoxia is usually observed after cardiac arrest as consequence of intubation and artificial ventilation with high fraction of oxygen (FiO₂) [19]. Mild hyperoxia can be defined as arterial oxygen partial pressure higher than 120 mmHg (> 16 kPa) and severe hyperoxia as PaO₂ > 200 mmHg (26.7 kPa). Elevated PaO₂ may increase oxidative stress, releases reactive oxygen species (ROS), and predisposes to mitochondrial dysfunction that finally results in molecular injury including deoxyribonucleic acid (DNA) damage and worsened organ function.

Elevated ribonucleic acid (RNA) expression of nitric oxide synthase (iNOS) was observed during hyperoxia when compared to normoxia in animal models of ischemia/reperfusion injury. Furthermore, pro apoptotic factors such as cyclooxygenase-2 (COX-2), toll like receptor 4 (TLR-4), and caspase-3 protein expression were increased, with a strong impact on apoptosis and inflammation mechanisms leading to myocardial injury [20].

Animal studies demonstrated improved outcome and lower neuronal damage using low O₂ supplementation compared with high O₂ supplementation. Recent data show an increased accumulation of hypoxic-induced factor (HIF) during exposure to oxygen FiO₂ > 30% in the brain tissue, hepatocytes, and myocardium [21–23].

Translated in the clinical practice, hyperoxia toxicity, with overproduction of ROS and oxidative cellular injury, can determinate elevated levels of superoxide, inhibits hypoxic pulmonary vasoconstriction, affects the adsorption of atelectasis, and is cause of apoptosis, necrosis, and direct ROS toxicity (Lorrain Smith effect) on alveolar capillary barrier which predisposes to hemorrhagic pulmonary edema; also, hyperoxia leads to vasoconstriction of arteries to distant organ thus causing coronary spasm and ischemic events [24].

Carbon dioxide

Youn-Jung Kim et al. [25] suggested that PaCO₂ may be a marker of post ROSC cerebral ischemia in nontraumatic cardiac arrest as PaCO₂ levels directly regulate cerebral blood flow and cerebrovascular diameter, with every 1 mmHg decrease of PaCO₂ determining a decrease of approximately 3% in cerebral blood flow. Hypocarbia may cause cerebral vasoconstriction that worsens ischemic injury and should be avoided to

preserve cerebral blood flow [26, 27]. On these basis, Eastwood et al. [28, 29] suggested that mild hypercapnia (PaCO₂ 50–55 mmHg) may improve cerebral oxygenation when compared with normocarbica (PaCO₂ 35–45 mmHg), and Beitler et al. [30] in a recent meta-analysis showed that neurocognitive outcome is improved by using low tidal volume ventilation. On the other hand, elevated levels of PaCO₂ can have detrimental effects on patients' outcome, especially if related to hypercapnic acidosis [31, 32].

Catecholamines

Catecholamines are principally used in cardiac arrest to achieve hemodynamic stabilization and to improve systemic blood pressure and cardiac output in hemodynamically unstable patients. However, recent studies showed that epinephrine administration is associated with post-resuscitation myocardial dysfunction and worse outcome [33]. Synthetic and endogenous catecholamines predispose to direct organ damage and cause many immunological, metabolic, and coagulation disturbances [34]. Catecholamines have “two faces”—are necessary to survive, but they can harm when in excess [35]. Adverse effects of catecholamines are connected to β₁ receptors binding, resulting in elevated myocardial oxygen consumption, myocardial failure, and left ventricular dysfunction. Raised levels of catecholamines predispose to β adrenoreceptors desensitization and downregulation of adrenoreceptors [36, 37]. Several conditions can increase plasma catecholamines up to toxic levels, including hypothyroidism, acidosis, and hypoxia as well as higher lactic acid [38].

Catecholamines can also affect peripheral vascular resistances through myosin light-chain kinase, which regulate smooth muscle cells myosin activity. The alpha 1 receptors activation induce vasoconstriction with increased risk of coronary spasms and can determinate reduction of splanchnic perfusion, lower mucosal pH, and higher hepatic vein lactate [39, 40]. Vasoconstriction, by increasing heart work, and ventricular overfilling predispose to ventricular arrhythmias.

Finally, catecholamine release, with consequent important alteration in microcirculatory system, impairs cardiac fatty-acid metabolism, and mitochondrial metabolism, and can activate immunology response. Neuronal and immune responses are rapidly activated during cardiac arrest and trigger a pathway of systemic neuroendocrine and immune responses with uncontrolled immune reaction because of inadequate hormonal suppression and disturbance between anti-inflammatory and proinflammatory responses. Catecholamines and their metabolites influence every aspect of innate and adaptive immune response, because immune cells express both alpha and beta adrenoreceptors [41]. In particular, catecholamines regulate T and B cells proliferation, differentiation, and apoptosis, and selectively activate lymphocytes subpopulations by release of pro- and anti-inflammatory mediators and are responsible for systemic activation and multiorgan damage [42].

The pathophysiology of the vicious circle after cardiac arrest

The above-described pathophysiological cross-talk between the immune system, catecholamine release, and metabolic status may result in the development of a dangerous pathophysiological vicious circle, where the ventilation and correction of arterial blood gases, as well as catecholamine administration may result in a paradox further

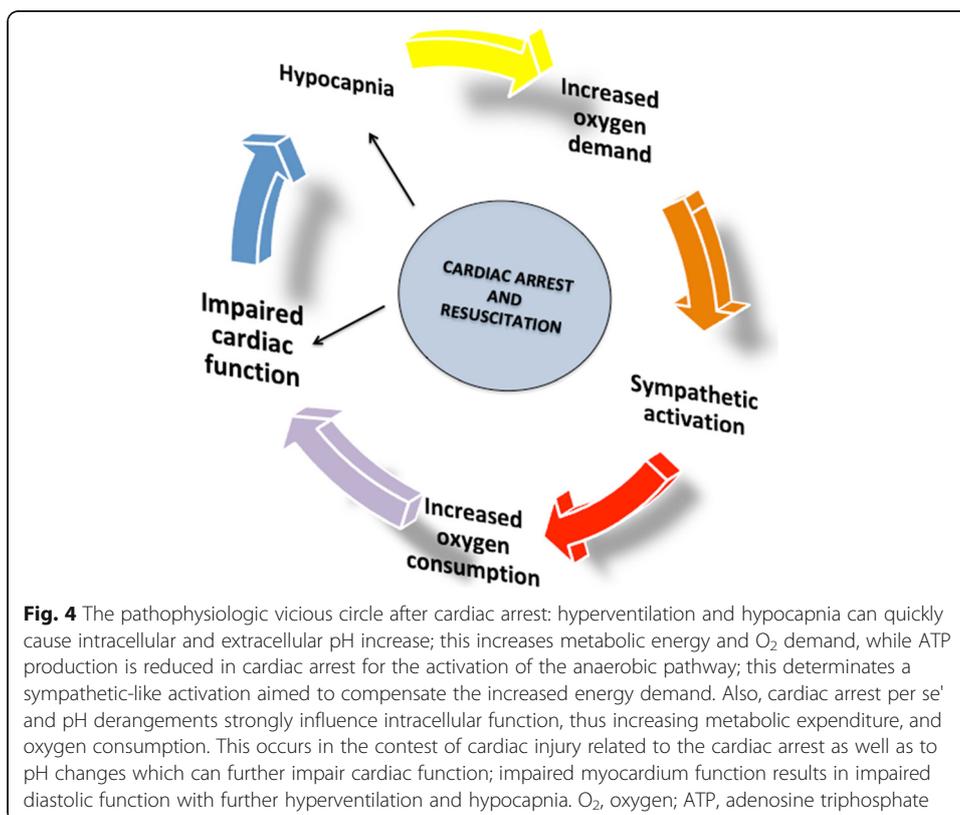
progression of primary damage. In particular, the pathophysiologic effects of carbon dioxide have a strong impact on the context of catecholamine administration and pH variations, but these effects are still poorly understood. CO₂ alterations during resuscitation can quickly cause abrupt intracellular and extracellular pH changes; hypocapnia, as consequence of hyperventilation, increases the demand of metabolic energy and O₂, while ATP production is often reduced, thus determining a sympathetic-like activation which further increases the energy demand (Fig. 4).

Metabolic compensatory mechanisms consequent to acute hypocapnia or hypercapnia are very slow, e.g., the renal metabolic compensation takes 5–7 days to restore the original intracellular pH [43].

The pH derangements strongly influence the function of many enzymes as well as the active and passive cytoplasmic transport of ions, especially of calcium (Ca²⁺) [44, 45]. Therefore, intracellular alkalosis during acute hypocapnia leads to increased metabolic expenditure, oxygen consumption, and increased metabolic demand [46]. On the other hand, pH changes can further impair cardiac function, which is common after cardiac arrest; impaired myocardium function results in worsened relaxation ability and increased diastolic pressure, resulting in increased pulmonary wedge pressure (PWP) and further hyperventilation and hypocapnia according to the above-described mechanisms [47].

A vicious circles can therefore occur, triggered by hypocapnia, which determinates a lack of energy substrate together with cardiac and pulmonary failure [48] (Fig. 4).

The increased production of catecholamines is a compensatory phenomenon in acidosis, and there is a positive feedback between catecholamine production and H⁺ concentration: the catecholamine-sensitivity is very low in acidosis, though their



production is high. However, when compensatory mechanisms and treatment improve acidosis and an alkalotic pH develops, the catecholamine levels remain still high; the synergism of the high levels of catecholamines and pH can poison systemic and immunological functions through the previously described mechanisms [47, 49].

Thus, the beneficial or harmful effect of catecholamines depends on different factors including pH, levels of PaCO₂, and lactate. In particular, the production of lactate depends on three mechanisms: hypoxia/anoxia, with activation of anaerobic pathway (lactate/pyruvate); ATP demand and rate of metabolism (low in acidosis and high in intracellular alkalosis), which is pH dependent; and intracellular pH [50]. Also, when tissue hypoxia is resolved, lactate production will terminate, and lactate degradation will start; therefore, the rapidity and the magnitude of the raise of carbon dioxide should ideally take into consideration the halftime of lactate degradation. Finally, lactate production is strictly determined by the pH of the intracellular space as it decreases in acidosis while it elevates in alkalosis.

Although from these pathophysiological concepts, it is clear that the best way to terminate this vicious circle is to elevate PaCO₂ levels, as acidosis can slow down the enzymatic intracellular and membrane processes by reducing energy demand—we should avoid raising PaCO₂ above normal level because this will lead to metabolic disorders in the long term [30].

Clinical implications

Post cardiac arrest syndrome results in different mechanisms of injury, including ischemia/reperfusion injury, oxidative stress, microcirculatory disturbances with coagulopathy, and inflammation which may increase the risk for multiple organ failure (Table 1).

During resuscitation, the relationship between tissue oxygenation, energy demand, and cellular function is complex and strictly related to intracellular pH.

Evidences regarding the levels and changes in oxygen and carbon dioxide during and after cardiac arrest as well as of optimal carbon dioxide and oxygenation targets are scarce, but the above discussed pathophysiological changes during cardiac arrest may help to guide clinical practice (Fig. 5).

During cardiac arrest, oxygen and carbon dioxide levels depend on the cause of pre arrest (respiratory or cardiological, hypothermia, etc.); two recent studies showed that during cardiopulmonary resuscitation, patients presenting with hypoxia had lower survival rates than those with normoxia or hyperoxia; in these studies, hypercapnia and respiratory acidosis were common, but no association between the severity of acidosis or hypercapnia with outcome was found [51, 52]. These results could be related to a quick decrease of carbon dioxide and pH normalization that could have harmful effects [53, 54].

Other authors showed that hyperventilation and hypocapnia during cardiopulmonary resuscitation can worsen outcome by reducing cardiac output and pulmonary blood flow [43, 44].

Therefore, current recommendations suggest to avoid hypoxia during cardiac arrest patients by using high FiO₂ 100% and a tidal volume of 500 ml, and respiratory rate of 12, minimizing acute changes on PaCO₂ [55] (Fig. 5).

In the post resuscitation phase, attention should be paid on FiO₂ administration, which should be titrated in order to avoid both hypoxia and hyperoxia. The use of high FiO₂ = 100% can cause extreme hyperoxia and can result in higher release of neuron specific enolase, a biomarker of brain injury, when compared to lower FiO₂ [46, 47].

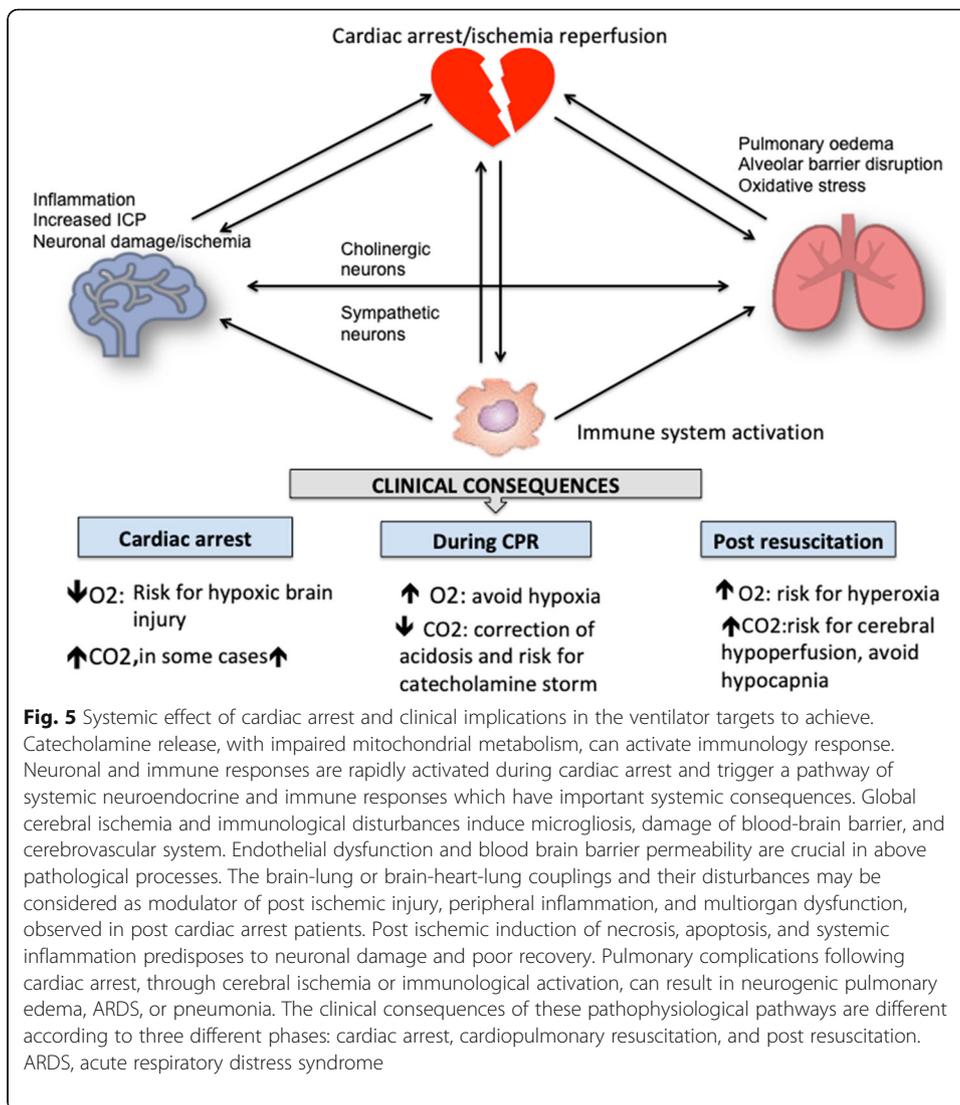
Table 1 Systemic consequences of cardiac arrest

	Pathophysiology	Clinical consequences
Ischemia-reperfusion injury	<ul style="list-style-type: none"> ✓ Free radical formation ✓ Apoptosis, necrosis ✓ "Sterile inflammation" ✓ Impaired resistance to infection ✓ Endothelial damage ✓ Impaired vasoregulation ✓ Adrenal suppression ✓ Electrolytes disturbances ✓ Acidosis, hypoxia, catabolism 	<ul style="list-style-type: none"> ✓ Multiorgan failure ✓ Infection/sepsis ✓ Microbiome disturbances ✓ Tissue hypoxia/ischemia ✓ Hyperglycemia ✓ Increase of intra-abdominal pressure ✓ Development of abdominal compartment syndrome
Brain injury/global cerebral ischemia (GCI)	<ul style="list-style-type: none"> ✓ Calcium homeostasis disruption ✓ No reflow ✓ Pyrexia ✓ Hyperglycemia ✓ Hyperoxygenation ✓ Impaired cerebrovascular autoregulation ✓ Impairment of hypothalamic-pituitary-adrenal axis ✓ Raised corticotropin releasing factor (CRF)-blood flow changes 	<ul style="list-style-type: none"> ✓ Damage of blood-brain barrier (BBB) (endothelial dysfunction and capillary permeability) ✓ Coma ✓ Seizures ✓ Cognitive dysfunction ✓ Stroke ✓ Brain death ✓ Pulmonary complications following cerebral ischemia (neurogenic pulmonary edema, acute distress respiratory syndrome, pneumonia)
Myocardial dysfunction	<ul style="list-style-type: none"> ✓ Global hypokinesia ✓ Preserved coronary blood flow ✓ Elevated systemic and pulmonary vascular resistance ✓ Impaired ventricular function ✓ Decreased venous return, cardiac compliance, and cardiac output. ✓ Decreased contractility 	<ul style="list-style-type: none"> ✓ Acute myocardial infarction (AMI) ✓ Arrhythmias ✓ Cardiomyopathies: dilated, restrictive, hypertrophic ✓ Cardiovascular collapse
Arrest related lung injury	<ul style="list-style-type: none"> ✓ Reperfusion injury ✓ Endothelial damage ✓ Systemic inflammation ✓ Activation of clotting cascades ✓ Ventilation-perfusion mismatch (ventilatory pressure) ✓ Basal atelectasis 	<ul style="list-style-type: none"> ✓ Pneumonia ✓ Acute respiratory distress syndrome ✓ Pulmonary thromboembolism
Arrest related liver and kidney injury	<ul style="list-style-type: none"> ✓ Reperfusion injury ✓ Reduced blood flow ✓ Reduced urine output 	<ul style="list-style-type: none"> ✓ Hypoxic hepatitis/acute liver failure ✓ Acute kidney injury

Peng et al. [20] in an experimental ischemia reperfusion injury model showed that hyperoxia after ischemia/reperfusion injury may be predictor of elevated oxidative stress, myocardial destruction, ventricular injury, and inflammation, compared to normoxia.

Ebner et al. [49] showed that both hyperoxemia or hypoxemia exposure after out hospital cardiac arrest are not associated with poor neurological outcome at 6 months, and that highest serum levels of neuronal injury marker -Tau (s-Tau) are not significantly correlated with PaO₂ values after 48 or 72 h from ROSC.

Hypocapnia should be avoided in order to optimize cerebral perfusion and mitigate ischemia-reperfusion injury due to post cardiac arrest oxidative stress, but the correction of hypocapnia should take in consideration the pH, catecholamine administration, and intracellular pH. In this phase, also the correction of hypercapnia should be performed with caution; some authors even suggest a correlation between good outcome and moderate hypercapnia by a potential effect of attenuation in the above mentioned excitotoxicity processes [50]. However, a recent study showed



that hypercapnia, especially when associated with acidosis, is associated with poor outcome, and the current recommendation is to aim for normoventilation with a PaO₂ of 4.5–6.0 kPa [31, 51](Fig. 4).

Patients after cardiac arrest also present with a variety of pulmonary complications, related to lung injury, aspiration, pulmonary ischemia-reperfusion injury, ventilator induced lung injury, and systemic inflammation and infections, which can affect oxygenation and ventilation.

Post cardiac arrest patients are at high risk for acute respiratory distress syndrome (ARDS), but the frequency of ARDS in this population is not well characterized. Two hypotheses have been proposed to explain the development of ARDS in these patients: the first theory includes the indirect activation and proliferation of neutrophils with progressive inflammatory alveolar infiltrates by activation of the immune system after cardiac arrest. Another hypothesis focuses on direct epithelial barrier damage related to ischemia reperfusion injury and endothelial cells activation and dysfunction [52]. The consequent overexpression of adhesion molecules, secretion of cytokines and

chemokines, activates the sequestration of neutrophils, which transmigrate across the endothelium and epithelium into the alveolar space and releases cytotoxic and proinflammatory compounds [52–55].

Gas exchange impairment after cardiac arrest can be also related to myocardial dysfunction and consequent pulmonary edema, as well as impairment of the immune system with increased predisposal to infections, with frequency up to 70% [56]. Finally, therapeutic hypothermia may increase the risk of infections [57]. In fact, post resuscitation syndrome presents characteristics of “sepsis like syndrome,” with high levels of cytokines and endotoxin in plasma which are similar to patients with sepsis and predispose to infections and longer mechanical ventilation [58] (Table 1).

Altogether, attention should be paid in the optimization of mechanical ventilation, in particular in the difference phases of cardiac arrest, during and in the post resuscitation period (Fig. 5), aiming to avoid hypo- and hyperoxia as well as maintain carbon dioxide levels and pH within physiological ranges.

Conclusions

The pathophysiological effects of cardiac arrest on systemic homeostasis are complex. The appropriate ventilator strategies are not fully understood, and oxygen and carbon dioxide targets should take in consideration several pathophysiological factors including the intracellular and extracellular pH as well as the use of catecholamines and hypothermia. In general, hypoxia (< 60 mmHg) and hyperoxia (> 300 mmHg) should be avoided as well as hypocapnia (< 30 mmHg) and hypercapnia (> 50 mmHg). Large prospective randomized controlled studies are warranted to further explore the pathophysiological basis of this theory and define the optimal setting of mechanical ventilation in patients after cardiac arrest.

Abbreviations

ARDS: Acute respiratory distress syndrome; ATP: Adenosine triphosphate; COX-2: Cyclooxygenase-2; DAMPS: Damage-associated molecular patterns; IL 1: Interleukin-1; iNOS: Nitric oxide synthase; MLCK: Myosin light-chain kinase; MODS: Multiple organ dysfunction syndrome; NO: Nitric oxide; ROS: Reactive oxygen species; ROSC: Return of spontaneous circulation; TLR 4: Toll like receptor; TNF: Tumor necrosis factor; VILI: Ventilator-induced lung injury

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References

1. Rellman AS (1972) Metabolic consequences of acid-base disorders. *Kidney Int.* 1:347–359
2. Kimmoun A, Novy E, Auchet T et al (2015) Hemodynamic consequences of severe lactic acidosis in shock states: from bench to bedside. *Crit Care* 19:175. <https://doi.org/10.1186/s13054-015-0896-7>
3. Phipers B, Pierce JT (2006) Lactate physiology in health and disease. *Contin Educ Anaesth Crit Care Pain* 6:128–132. <https://doi.org/10.1093/bjaceaccp/mkl018>
4. Levy B (2006) Lactate and shock state: the metabolic view. *Curr Opin Crit Care* 12:315–321. <https://doi.org/10.1097/01.ccx.0000235208.77450.15>
5. Wutrich Y, Barraud D, Conrad M et al (2010) Early increase in arterial lactate concentration under epinephrine infusion is associated with a better prognosis during shock. *Shock* 34:4–9. <https://doi.org/10.1097/SHK.0b013e3181ce2d23>
6. Sanders AB, Otto CW, Kern KB et al (1988) Acid-base balance in a canine model of cardiac arrest. *Ann Emerg Med* 17:667–671
7. Ellison GM, Torella D, Karakikes I et al (2007) Acute beta-adrenergic overload produces myocyte damage through calcium leakage from the ryanodine receptor 2 but spares cardiac stem cells. *J. Biolumin. Chemilumin.* 282:11397–11409. <https://doi.org/10.1074/jbc.M607391200>
8. Teplinsky K, O'Toole M, Olman M et al (1990) Effect of lactic acidosis on canine hemodynamics and left ventricular function. *Am J Physiol Circ Physiol* 258:H1193–H1199. <https://doi.org/10.1152/ajpheart.1990.258.4.H1193>
9. Marsh JD, Margolis TI, Kim D (1988) Mechanism of diminished contractile response to catecholamines during acidosis. *Am J Physiol* 254:H20–H27. <https://doi.org/10.1152/ajpheart.1988.254.1.H20>
10. Rajamäki K, Nordström T, Nurmi K et al (2013) Extracellular acidosis is a novel danger signal alerting innate immunity via the NLRP3 inflammasome. *J. Biolumin. Chemilumin.* 288:13410–13419. <https://doi.org/10.1074/jbc.M112.426254>
11. Mendler AN, Hu B, Prinz PU et al (2012) Tumor lactic acidosis suppresses CTL function by inhibition of p38 and JNK/c-Jun activation. *Int. J. Cancer* 131:633–640. <https://doi.org/10.1002/ijc.26410>
12. Martínez D, Vermeulen M, von Ew E et al (2007) Extracellular acidosis triggers the maturation of human dendritic cells and the production of IL-12. *J. Immunol.* 179:1950–1959
13. Pedoto A, Nandi J, Oler A et al (2001) Role of nitric oxide in acidosis-induced intestinal injury in anesthetized rats. *J Lab Clin Med* 138:270–276. <https://doi.org/10.1067/mlc.2001.118176>
14. Morrison LJ, Deakin CD, Morley PT et al (2010) Part 8: advanced life support: 2010 international consensus on cardiopulmonary resuscitation and emergency cardiovascular care science with treatment recommendations. *Circulation* 122:S345–S421. <https://doi.org/10.1161/CIRCULATIONAHA.110.971051>
15. Helmerhorst HJF, Roos-Blom M-J, Van Westerloo DJ, et al (2015) Associations of arterial carbon dioxide and arterial oxygen concentrations with hospital mortality after resuscitation from cardiac arrest. doi: <https://doi.org/10.1186/s13054-015-1067-6>
16. Elmer J, Wang B, Melhem S et al (2015) Exposure to high concentrations of inspired oxygen does not worsen lung injury after cardiac arrest. *Crit Care* 19:105. <https://doi.org/10.1186/s13054-015-0824-x>
17. Michiels C (2004) Physiological and pathological responses to hypoxia. *Am J Pathol* 164:1875–1882. [https://doi.org/10.1016/S0002-9440\(10\)63747-9](https://doi.org/10.1016/S0002-9440(10)63747-9)
18. Pope JV, Jones AE, Gaieski DF et al (2010) Multicenter study of central venous oxygen saturation (ScvO₂) as a predictor of mortality in patients with sepsis. *Ann Emerg Med* 55:40–46.e1. <https://doi.org/10.1016/j.annemergmed.2009.08.014>
19. Harmon MBA, van Meenen DMP, van der Veen ALIP et al (2018) Practice of mechanical ventilation in cardiac arrest patients and effects of targeted temperature management: a substudy of the targeted temperature management trial. *Resuscitation* 129:29–36. <https://doi.org/10.1016/j.resuscitation.2018.04.040>
20. Peng Y-W, Mohammed A, Deatrick KB et al (2018) Differential effects of normoxic and hyperoxic reperfusion on global myocardial ischemia-reperfusion injury. *Semin Thorac Cardiovasc Surg.* <https://doi.org/10.1053/J.SEMTCVS.2018.09.018>
21. Richards EM, Fiskum G, Rosenthal RE et al (2007) Hyperoxic reperfusion after global ischemia decreases hippocampal energy metabolism. *Stroke* 38:1578–1584. <https://doi.org/10.1161/STROKEAHA.106.473967>
22. Balan IS, Fiskum G, Hazelton J et al (2006) Oximetry-guided reoxygenation improves neurological outcome after experimental cardiac arrest. *Stroke* 37:3008–3013. <https://doi.org/10.1161/01.STR.0000248455.73785.b1>
23. Keeley TP, Mann GE (2019) Defining physiological normoxia for improved translation of cell physiology to animal models and humans. *Phys. Rev.* 99:161–234. <https://doi.org/10.1152/physrev.00041.2017>
24. Heinrichs J, Lodewyckx C, Neilson C et al (2018) The impact of hyperoxia on outcomes after cardiac surgery: a systematic review and narrative synthesis. *Can J Anesth Can d'anesthésie* 65:923–935. <https://doi.org/10.1007/s12630-018-1143-x>
25. Kim Y-J, Lee YJ, Ryoo SM, Sohn CH, Ahn S, Seo D-W, Lim KS, Kim WY (2016) Role of blood gas analysis during cardiopulmonary resuscitation in out-of-hospital cardiac arrest patients. *Medicine (Baltimore)* 95:e3960. <https://doi.org/10.1097/md.0000000000003960>
26. Willie CK, Tzeng Y-C, Fisher JA, Ainslie PN (2014) Integrative regulation of human brain blood flow. *J. Phys.* 1592:841–859. <https://doi.org/10.1113/jphysiol.2013.268953>
27. Tejerina E, Pelosi P, Muriel A et al (2017) Association between ventilatory settings and development of acute respiratory distress syndrome in mechanically ventilated patients due to brain injury. *J Crit Care* 38:341–345. <https://doi.org/10.1016/j.jcrc.2016.11.010>

28. Eastwood GM, Young PJ, Bellomo R (2014) The impact of oxygen and carbon dioxide management on outcome after cardiac arrest. *Curr Opin Crit Care* 20:266–272. <https://doi.org/10.1097/MCC.0000000000000084>
29. Eastwood GM, Nichol A, Wise MP. Targeted therapeutic mild hypercapnia after cardiac arrest. doi: <https://doi.org/10.1186/s13054-017-1670-9>
30. Beitler JR, Ghafoori TB, Jinadasa SP et al (2017) Favorable neurocognitive outcome with low tidal volume ventilation after cardiac arrest. *Am J Respir Crit Care Med* 195:1198–1206. <https://doi.org/10.1164/rccm.201609-1771OC>
31. Tiruvoipati R, Pilcher D, Botha J et al (2018) Association of hypercapnia and hypercapnic acidosis with clinical outcomes in mechanically ventilated patients with cerebral injury. *JAMA Neurol* 75:818. <https://doi.org/10.1001/jamaneurol.2018.0123>
32. Ebner F, Harmon MBA, Aneman A et al (2018) Carbon dioxide dynamics in relation to neurological outcome in resuscitated out-of-hospital cardiac arrest patients: an exploratory Target Temperature Management Trial substudy. *Crit Care* 22:196. <https://doi.org/10.1186/s13054-018-2119-5>
33. Mauch J, Ringer S, Spielmann N, Weiss M (2014) Impact of catecholamines in cardiac arrest due to acute asphyxia—a study in piglets. *Pediatr Anesth* 24:933–939. <https://doi.org/10.1111/pan.12457>
34. O'Connor CM, Gattis WA, Uretsky BF et al (1999) Continuous intravenous dobutamine is associated with an increased risk of death in patients with advanced heart failure: insights from the Flolan International Randomized Survival Trial (FIRST). *Am Heart J* 138:78–86
35. Perkins GD, Ji C, Deakin CD et al (2018) A randomized trial of epinephrine in out-of-hospital cardiac arrest. *N Engl J Med* 379:711–721. <https://doi.org/10.1056/NEJMoa1806842>
36. Bristow MR (1998) Why does the myocardium fail? Insights from basic science. *Lancet* (London, England) 352 Suppl:S18–14. doi: [https://doi.org/10.1016/S0140-6736\(98\)90311-7](https://doi.org/10.1016/S0140-6736(98)90311-7)
37. Wang W, Hua T, Li H et al (2018) Decreased cAMP level and decreased downregulation of β_1 -adrenoceptor expression in therapeutic hypothermia-resuscitated myocardium are associated with improved post-resuscitation myocardial function. *J Am Heart Assoc*. <https://doi.org/10.1161/JAHA.117.006573>
38. Rivers EP, Wortsman J, Rady MY et al (1994) The effect of the total cumulative epinephrine dose administered during human CPR on hemodynamic, oxygen transport, and utilization variables in the postresuscitation period. *Chest* 106:1499–1507
39. Xiang Y, Kobilka BK (2003) Myocyte adrenoceptor signaling pathways. *Science* (80-) 300:1530–1532. <https://doi.org/10.1126/science.1079206>
40. Meier-Hellmann A, Reinhart K, Bredle DL et al (1997) Epinephrine impairs splanchnic perfusion in septic shock. *Crit Care Med* 25:399–404
41. Sanders VM (2012) The beta2-adrenergic receptor on T and B lymphocytes: do we understand it yet? *Brain Behav Immun* 26:195–200. <https://doi.org/10.1016/j.bbi.2011.08.001>
42. Bergquist J, Tarkowski A, Ekman R, Ewing A (1994) Discovery of endogenous catecholamines in lymphocytes and evidence for catecholamine regulation of lymphocyte function via an autocrine loop. *Proc Natl Acad Sci U S A*. 91:12912–12916
43. Aufderheide TP, Sigurdsson G, Pirralo RG et al (2004) Hyperventilation-induced hypotension during cardiopulmonary resuscitation. *Circulation* 109:1960–1965. <https://doi.org/10.1161/01.CIR.0000126594.79136.61>
44. Yannopoulos D, Tang W, Roussos C et al (2005) Reducing ventilation frequency during cardiopulmonary resuscitation in a porcine model of cardiac arrest. *Respir Care* 50:628–635
45. Sutherasan Y, Raimondo P, Pelosi P (2015) Ventilation and gas exchange management after cardiac arrest. *Best Pract Res Clin Anaesthesiol* 29:413–424. <https://doi.org/10.1016/j.bpa.2015.09.001>
46. Nelskylä A, Parr MJ, Skrifvars MB (2013) Prevalence and factors correlating with hyperoxia exposure following cardiac arrest—an observational single centre study. *Scand J Trauma Resusc Emerg Med* 21:35. <https://doi.org/10.1186/1757-7241-21-35>
47. Kuisma M, Boyd J, Voipio V et al (2006) Comparison of 30 and the 100% inspired oxygen concentrations during early post-resuscitation period: a randomised controlled pilot study. *Resuscitation* 69:199–206. <https://doi.org/10.1016/j.resuscitation.2005.08.010>
48. Sikter A (2018) Hypocapnia and mental stress can trigger vicious circles in critically ill patients due to energy imbalance: a hypothesis presented through cardiogenic pulmonary oedema. *Neuropsychopharmacol Hung* 20:65–74
49. Ebner F, Ullén S, Aneman A et al (2019) Associations between partial pressure of oxygen and neurological outcome in out-of-hospital cardiac arrest patients: an explorative analysis of a randomized trial. *Crit Care* 23:30. <https://doi.org/10.1186/s13054-019-2322-z>
50. Vaahersalo J, Bendel S, Reinikainen M et al (2014) Arterial blood gas tensions after resuscitation from out-of-hospital cardiac arrest. *Crit Care Med* 42:1463–1470. <https://doi.org/10.1097/CCM.0000000000000228>
51. Soar J, Nolan JP, Böttiger BW et al (2015) European resuscitation council guidelines for resuscitation 2015. *Resuscitation* 95:100–147. <https://doi.org/10.1016/j.resuscitation.2015.07.016>
52. Johnson NJ, Carlbom DJ, Gaieski DF (2018) Ventilator management and respiratory care after cardiac arrest. *Chest* 153: 1466–1477. <https://doi.org/10.1016/j.chest.2017.11.012>
53. Spindelboeck W, Gemes G, Strasser C et al (2016) Arterial blood gases during and their dynamic changes after cardiopulmonary resuscitation: a prospective clinical study. *Resuscitation* 106:24–29. <https://doi.org/10.1016/j.resuscitation.2016.06.013>
54. Spindelboeck W, Schindler O, Moser A et al (2013) Increasing arterial oxygen partial pressure during cardiopulmonary resuscitation is associated with improved rates of hospital admission. *Resuscitation* 84:770–775. <https://doi.org/10.1016/j.resuscitation.2013.01.012>
55. Sutherasan Y, Varghas M, Brunetti I, Pelosi P (2015) Ventilatory targets after cardiac arrest. *Minerva Anesthesiol* 81:39–51
56. Mongardon N, Perbet S, Lemiale V et al (2011) Infectious complications in out-of-hospital cardiac arrest patients in the therapeutic hypothermia era. *Crit Care Med* 39:1359–1364. <https://doi.org/10.1097/CCM.0b013e3182120b56>
57. Sagalyn E, Band RA, Gaieski DF, Abella BS (2009) Therapeutic hypothermia after cardiac arrest in clinical practice: review and compilation of recent experiences. *Crit Care Med* 37:S223–S226. <https://doi.org/10.1097/CCM.0b013e3181aa5c7c>
58. Sutherasan Y, Peñuelas O, Muriel A et al (2015) Management and outcome of mechanically ventilated patients after cardiac arrest. *Crit Care* 19:215. <https://doi.org/10.1186/s13054-015-0922-9>

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