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# Liver–lung interactions in acute respiratory distress syndrome



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## Abstract

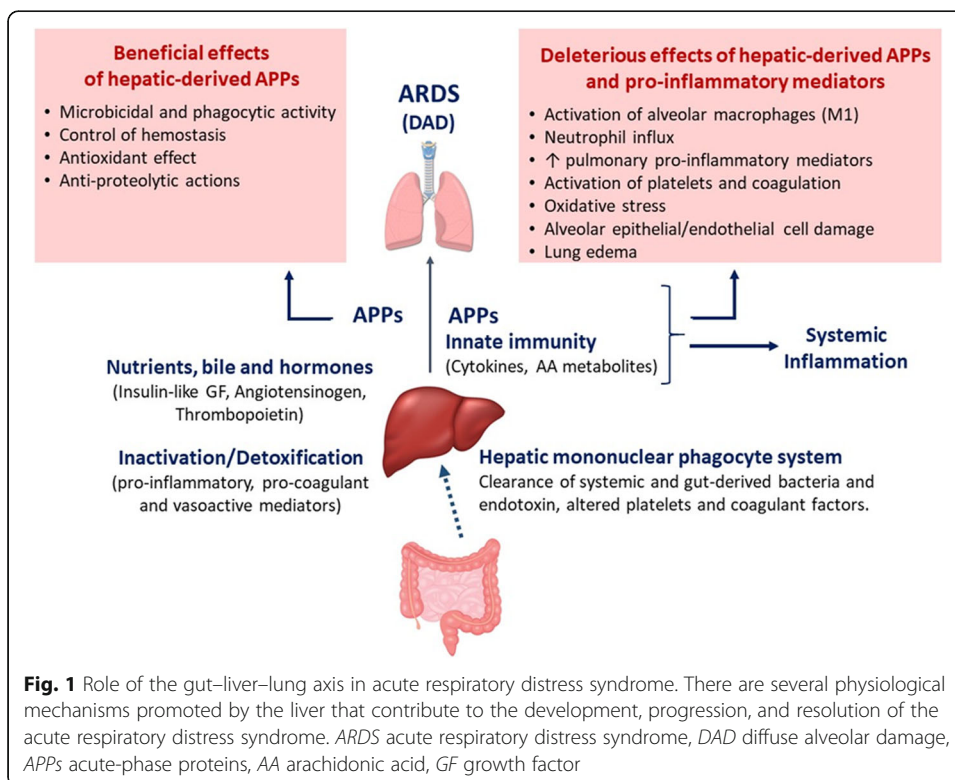
Patients with liver diseases are at high risk for the development of acute respiratory distress syndrome (ARDS). The liver is an important organ that regulates a complex network of mediators and modulates organ interactions during inflammatory disorders. Liver function is increasingly recognized as a critical determinant of the pathogenesis and resolution of ARDS, significantly influencing the prognosis of these patients. The liver plays a central role in the synthesis of proteins, metabolism of toxins and drugs, and in the modulation of immunity and host defense. However, the tools for assessing liver function are limited in the clinical setting, and patients with liver diseases are frequently excluded from clinical studies of ARDS. Therefore, the mechanisms by which the liver participates in the pathogenesis of acute lung injury are not totally understood. Several functions of the liver, including endotoxin and bacterial clearance, release and clearance of pro-inflammatory cytokines and eicosanoids, and synthesis of acute-phase proteins can modulate lung injury in the setting of sepsis and other severe inflammatory diseases. In this review, we summarized clinical and experimental support for the notion that the liver critically regulates systemic and pulmonary responses following inflammatory insults. Although promoting inflammation can be detrimental in the context of acute lung injury, the liver response to an inflammatory insult is also pro-defense and pro-survival. A better understanding of the liver–lung axis will provide valuable insights into new diagnostic targets and therapeutic strategies for clinical intervention in patients with or at risk for ARDS.

**Keywords:** Liver–lung interaction, Acute respiratory distress syndrome, Liver dysfunction, Mechanisms, Immunomodulation, Acute-phase response, Critical illness

### Background

Acute respiratory distress syndrome (ARDS) is a severe respiratory failure, due to non-cardiogenic pulmonary edema [1, 2], associated with a hospital mortality between 35% and 46% [1, 3, 4]. The pathology of ARDS involves diffuse alveolar damage (DAD), which comprises severe alveolar epithelial cell damage, neutrophil infiltration, activation of alveolar macrophages, production of cytokines and chemokines, plasma extravasation, procoagulant activity with fibrin deposition, hyaline membrane formation, myofibroblast proliferation, and fibrosis in the intra-alveolar spaces [2, 4]. Formation of protein-rich edema in the airspaces due to the disruption of the alveolar–capillary membrane is one of the main factors that contributes to the severe impairment of blood and tissue oxygenation early in the evolution of DAD [2, 4]. The DAD occurs not only in response to a direct injury to the lung (e.g., pneumonia), but it may also represent a pulmonary manifestation of diverse systemic immunoregulatory disorders, such as sepsis [4]. The pathogenesis of ARDS, therefore, is linked to changes in local and systemic host defense and immune responses [5], in which the liver plays an important role (Fig. 1).

The liver has unique anatomic, cellular, and physiological characteristics that enable the clearance of circulating microbial products, tissue debris, altered platelets, products of intravascular coagulation, and different bioactive molecules (Fig. 1) [6–10]. Also, the liver has a key role in the synthesis of proteins, metabolism of toxins and drugs, and in the modulation of systemic inflammatory responses and host defense (Fig. 1). It is becoming more evident that normal liver function exerts lung protection and is necessary for recovery from lung damage [11, 12]. In this line, it has been observed that established ARDS during acute liver allograft rejection is resolved within hours of



hepatic re-transplantation [13]. On the other hand, experimental studies suggest that the presence of the liver is also absolutely necessary for inducing lung injury in rats [14]. These apparently paradoxical observations highlight the relevant crosstalk between lung and liver in ARDS.

Despite the well-recognized liver–lung interaction in the pathogenesis of ARDS, its underlying mechanisms and its effects on the outcome of these patients have been barely studied due to several reasons. First, patients with liver diseases are frequently excluded from studies of ARDS. In addition, liver function is not precisely reflected by the standard liver function tests in the clinical setting, and the liver is not as accessible as other organs such as the lung, making liver dysfunction not as evident as dysfunctions of other organs. Finally, its clinical consequences are also heterogeneous in critically ill patients [15]. The present work reviews the important role of the liver on the development and resolution of ARDS and aims to provide an integrated view of the underlying mechanisms that support the liver–lung interaction in critically ill patients.

### **The reciprocal impacts of lung and liver dysfunctions**

Following hepatocellular damage, the liver may reduce its clearance function, increase the synthesis of deleterious substances, and dysregulate immune responses, leading to systemic complications such as coagulopathy, elevated risk of infection, hypoglycaemia, exacerbated inflammatory responses, encephalopathy, and damage of other extrahepatic organs, including lung injury [16–22]. In critically ill patients, hepatic dysfunction is recognized as a relevant clinical condition that influences the development, severity, and progression of ARDS [5, 11, 19, 23–27]. In ARDS patients, liver dysfunction is a major determinant of mortality [24–26]. It is well known that cirrhosis and other chronic liver diseases make the patients more susceptible for developing ARDS, which adversely affects patient outcomes [24–26, 28]. A growing body of evidence suggests that liver damage activates and enhances inflammation in the pulmonary intravascular compartment and lower respiratory tract, leading to important changes in the structure and/or functions of the lung [29, 30]. Although all these observations indicate that liver function is an important factor for the development and resolution of ARDS, there is also evidence that such interorgan communication is bidirectional. Thus, acute lung injury is known to impair hepatic function and to aggravate liver diseases by mechanisms involving hypoxemia, activation of systemic inflammatory responses, and cardiovascular changes [24, 31, 32].

### **Liver dysfunction is common in critical care patients**

The frequency of liver damage in critical illness has considerably increased over the last decades [23, 33–35], reaching up to 20% of ICU patients in some series and elevating their morbi-mortality [33, 34, 36]. In critically ill patients, liver dysfunction usually occurs after inflammatory insults such as sepsis and trauma [15, 23, 33, 37], and the underlying interactive mechanisms are complex. The mechanisms of liver dysfunction in critically ill patients implicate microbial products, the paracrine action of cytokines and other inflammatory mediators, hypoxemia, oxidative stress, toxic compounds, hypoperfusion, passive congestion, and effect of nutrition support, among others [34, 38–40].

Liver dysfunction can be manifested by plasma elevation of liver enzymes (aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP),  $\gamma$ -glutamyl transpeptidase) and bilirubin, decreased plasma levels of albumin and coagulation factors, and/or increased international normalized ratio (INR) [33, 36]. The clearance rate of indocyanine green has been used as a dynamic test to assess the functional capacity of the liver. Although the indocyanine green test has shown to reflect better the excretory and/or microvascular dysfunction of the liver, its clinical use has certain limitations [41]. Increased plasma levels of bilirubin are associated with high mortality in critically ill patients [39, 42]. Furthermore, hyperbilirubinemia has been proposed as a biomarker of ARDS and found to be an independent factor of mortality in patients with ARDS [26, 35, 39, 43]. Unfortunately, neither bilirubinemia nor other hepatic parameters routinely measured in the clinical setting have the sensitivity and specificity required for an early identification of hepatic injury in critically ill patients [23, 33, 36, 40].

### **Mechanisms of liver–lung interactions in ARDS**

The mechanisms by which the liver modulates lung injury involve interrelated elements of systemic and pulmonary host defense, inflammatory responses, and metabolism and include the following (see Fig. 1).

#### **Clearance by the hepatic mononuclear phagocyte system of systemic endotoxemia, bacteremia, vasoactive by-products, and procoagulant factors**

The mononuclear phagocyte system located in the liver, spleen, lung, and bone marrow constitutes the major mechanism to uptake and detoxify bacteria, fungi, viruses, and dying cells, limiting the magnitude and duration of infections [10, 44, 45]. Although these mononuclear phagocytic cells can exert this function in all these locations, their major mass is in the hepatic sinusoids [10, 46]. The hepatic sinusoid is a unique vascular structure with highly specialized endothelial cells (liver sinusoidal endothelial cells) and liver macrophages (Kupffer cells) that reside within the lumen. The cells of the hepatic sinusoid are constantly exposed to gut-derived bacteria, microbial debris, and bacterial endotoxins. Kupffer cells, which line the extensive sinusoidal network, constitute nearly 80–90% of the tissue macrophages present in the body and exert an important role in host defense through phagocytosis and a multitude of secretory functions [46]. The hepatic mononuclear phagocyte system acts as a first line of defense in clearing bacteria and their products. Besides uptake of microbial pathogens and products, Kupffer cells also protect the lung and other extrahepatic organs by removing altered platelets and intravascular coagulation products (Fig. 1) [8–10, 46].

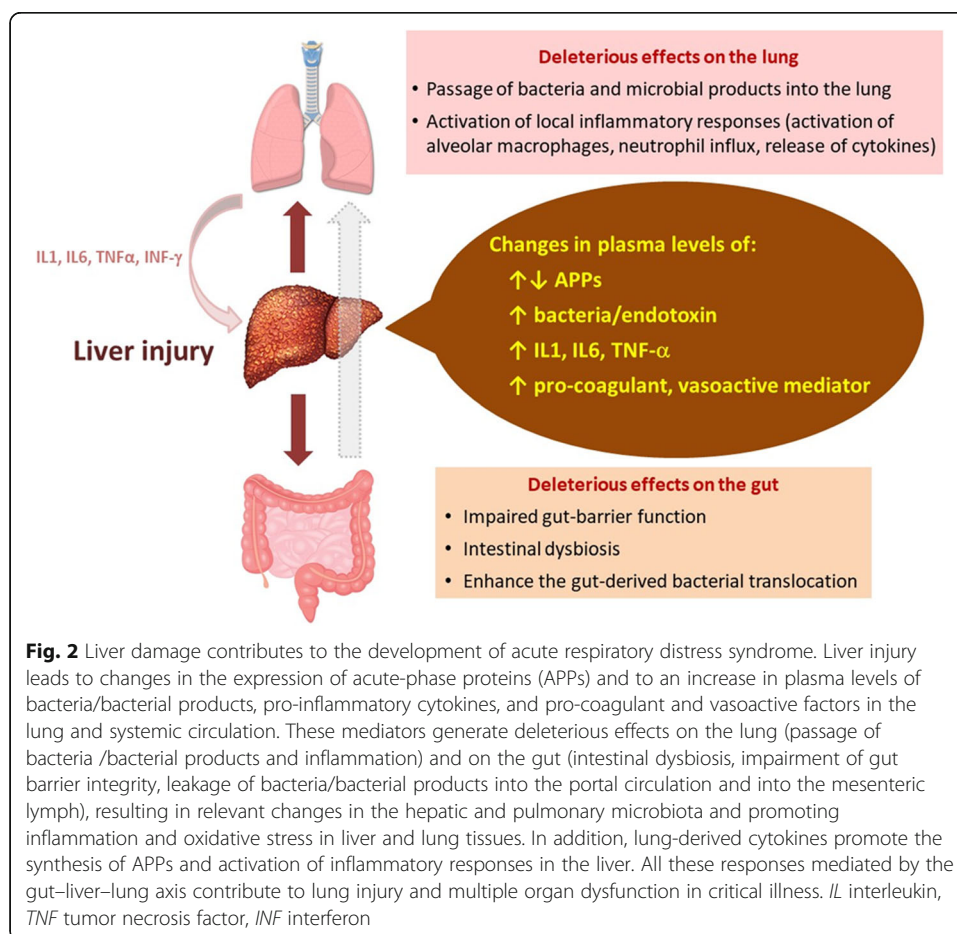
Dysfunction of the reticuloendothelial system of the liver allows bacterial and microbial products, including the so-called pathogen-associated molecular patterns (PAMPs) [47], to reach the lung and the systemic circulation, where they activate pulmonary and systemic inflammatory responses (Fig. 1) [29, 47–49]. Indeed, increased plasma levels of endotoxin, probably of intestinal origin, along with increased levels of some cytokines have been found in the blood of patients with acute and chronic liver diseases [50–54].

Pulmonary deposition of intravascular bacteria, and their products alter the structure and function of the lung by different mechanisms including (i) direct cytotoxic effect

on alveolar epithelial and endothelial cells, (ii) modulation of local innate immune responses in the lung via activation of toll-like receptors (TLRs), resulting in activation of resident alveolar macrophages and neutrophil influx and in the production of reactive oxygen species, (iii) activation of the coagulation cascades and platelet aggregation, leading to pulmonary microvascular thrombosis [29, 48, 49, 55–59], and (iv) a sustainable increase in pulmonary vascular resistance [22, 30] (Fig. 2). All these mechanisms alter the alveolar endothelial and epithelial cell functions and enhance barrier permeability leading to the formation pulmonary alveolar edema and respiratory failure [14, 60], the two main characteristics of ARDS (Fig. 2).

### Metabolic inactivation and detoxification of endogenous inflammatory mediators

The hepatobiliary system has an important capacity to inactivate and detoxicate pro-inflammatory cytokines, vasoactive mediators, and eicosanoids from the systemic circulation. Removal of all these mediators constitutes a critical element of systemic and pulmonary host defense, protecting the lung and other extrahepatic organs from injury (Fig. 1) [8–10, 46]. Like endotoxin, increased levels of cytokines (such as IL-8, IL-1 $\beta$ , ENA-78, TNF- $\alpha$ , MCP-1, MIP-1 $\alpha$ ,...) and arachidonic acid-derived eicosanoids (thromboxane, leukotrienes) not cleared by the liver have been shown to exert a direct cytotoxic effect on alveolar epithelial and endothelial cells, to activate local innate



immune responses and to promote platelet aggregation in the lung, contributing to the development of diffuse alveolar damage (DAD) [55–59].

#### **Hepatic synthesis of inflammatory mediators that can activate pulmonary alveolar macrophages and, consequently, increase inflammation in the lung**

Hepatic mononuclear cells include a heterogeneous population of lymphocytes, Kupffer cells (hepatic resident macrophages), monocytes, and granulocytes that perform vital functions for the innate and adaptive immune system. In response to injury, activation of these hepatic mononuclear cells enhances the production and release of inflammatory mediators, such as IL-1, IL-6, TNF- $\alpha$ , platelet-activating factor (PAF), and leukotrienes, into the systemic circulation [61], where they play an important role in the lung–liver interaction [18, 31, 51, 61–64]. These liver-derived inflammatory mediators alter lung structure function early in acute inflammatory diseases (such as sepsis) and contribute to some extent to lung damage upon activation of pulmonary alveolar macrophages (Fig. 1) [17, 65]. In this line, elevated levels of TNF- $\alpha$  and IL-1 $\beta$ , two cytokines that are mainly synthesized by alveolar macrophages, have been found in the lungs of rats with carbon tetrachloride (CCl<sub>4</sub>)-induced cirrhosis, along with an increase in lipid peroxidation (TBARS) and antioxidant enzymes (superoxide dismutase and catalase) in the liver and lung tissues. These events are also associated with altered gas exchange and changes in the size of pulmonary vessels in these rats [66, 67]. Besides high levels of endotoxin [50, 68], patients with liver disorders also have high circulating levels of TNF- $\alpha$ , IL-1, and IL-6 [51–54] because of the altered capacity for inactivation and detoxification and the increased synthesis of pro-inflammatory mediators by the liver [9, 44, 46, 61]. These specific cytokines have been shown to modulate systemic inflammatory responses and participate in the development of lung damage [69–72]. Therefore, it is possible that cytokines of hepatic origin may control and modulate the local host defense and immune system of the lung, contributing to lung injury (Figure 2).

#### **The liver is the main organ responsible for the acute-phase response**

The organism responds to tissue injury or infection by local changes such as those associated to inflammation and by a coordinated sequence of systemic and metabolic process, known as the acute-phase response, aimed to restore homeostasis and recover from injury [63, 73–76]. One of the major characteristics of this acute-phase response is a change in plasma concentration (either increase or decrease) of the acute-phase proteins (APPs) expressed in the liver [74]. Cytokine-driving synthesis of acute-phase proteins in the liver modulates the systemic and pulmonary host inflammatory responses and intermediary metabolism (Fig. 1) [22, 30, 48]. The hepatic APPs have a variety of functions that include microbicidal and phagocytic activity (e.g., LPS binding protein, complement components, C-reactive protein), recruitment of immune cells to inflammatory sites (e.g., serum amyloid A), hemostasis (e.g., fibrinogen,  $\alpha$ 1-acid glycoprotein), antioxidant, and prevention of iron loss (e.g., haptoglobin) and anti-proteolytic actions to counterbalance protease activity at sites of inflammation (e.g.,  $\alpha$ 2-macroglobulin,  $\alpha$ 1-antitrypsin, and  $\alpha$ 1-antichymotrypsin) (Fig. 1) [63, 74, 75].



While the local inflammation occurs in the alveolar airspaces of patients with ARDS, the acute-phase response is induced in the liver [73, 76, 77]. Interestingly, in pneumonia-induced ARDS, this liver-derived acute-phase response occurs independently of bacterial dissemination and depends instead on inflammatory signaling molecules derived from the pulmonary immune cells, such as the cytokines IL-1, IL-6, and TNF- $\alpha$  [10, 62, 63, 73, 74]. Then, these lung-derived cytokines can travel from the lung into the systemic circulation and ultimately modify acute-phase gene expression in the liver [63, 73, 78, 79] upon activation of the transcription factors STAT3 (signal transducer and activator of transcription 3) by IL-6 and activation of RelA (v-rel avian reticuloendotheliosis viral oncogene homolog A, also known as NF- $\kappa$ B3) by the early-response cytokines TNF- $\alpha$  and IL-1 [73, 78, 79]. In response to these cytokines, the liver changes the expression of many acute-phase proteins such as C-reactive protein,  $\alpha$ -1 antitrypsin, serum amyloid A protein, and others [10, 62, 63, 74, 77], which in turn can directly travel back to the lung and pass into the airspaces where they cause inflammation, predominantly via activation of alveolar macrophages (Fig. 2) [5, 14]. These phagocytic cells are targeted by multiple hepatic APPs such as SAA [80, 81], SAP [82], LBP [83], and C-reactive protein [84, 85]. Once activated by these hepatic APPs, alveolar macrophages release cytokines (IL-6 and CXCL1) that enhance local inflammation, in part by promoting neutrophil influx to the interstitium and alveolar airspaces. Excessive inflammation in the alveoli may result in an increase in oxidative stress and lung injury [86, 87]. Besides this potentially deleterious effect, hepatic APPs at the site of plasma extravasation has other potential functions, including opsonization of bacteria, leukocyte activation, antiprotease, antioxidant activities, and modulation of the coagulation pathway [63, 75]. These mechanisms help to regulate host defense, limit excessive inflammation and immune responses, and promote bacterial clearance, preventing infection dissemination and reducing the risk of organ damage in the setting of pneumonia and sepsis. Also, hepatic APPs exert liver protection by countering TNF-dependent toxicity in the liver and attenuate systemic inflammation and mortality in sepsis and pneumonia-induced ARDS (Fig. 1) [79, 86, 88–90]. Altogether, the bidirectional liver–lung axis mediated by APPs is critical for integrating systemic and pulmonary responses, balancing regulation of multiple host defenses and activation of inflammation to restore homeostasis and recover from organ injury [48, 61, 86]. Disbalance in this liver–lung communication can be an important factor in the initiation and progression of ARDS and of the damage to other organs [73].

#### **Nutrients, bile, and hormone production**

The liver plays an important role in regulating metabolic homeostasis and in the synthesis and processing of lipids and carbohydrates that supply energy to other organs [91]. It is also the major site of synthesis of key proteins and bile acids that are critical for the normal uptake of vitamins and lipids [92]. Therefore, alterations in the flux of carbohydrates and lipids through the liver can indirectly impact distal organs due to alteration of their energy statuses [93]. In addition, hyperbilirubinemia in the context of liver diseases has been shown to cause some lung-specific deleterious effects, by entering the lung tissue, reaching the alveolar airspaces, and deteriorating the surface

tension properties of the alveolar surfactant [94]. Although bilirubin has antioxidant properties, high bilirubin levels can also activate oxidative stress, apoptosis, and inflammatory responses in different cell types and organs [95–98]. Therefore, hyperbilirubinaemia may actively participate in the development of ARDS, although the underlying mechanisms have not been fully elucidated. Finally, the liver produces several hormones that mediate diverse extrahepatic effects, such as insulin-like growth factor, angiotensinogen, and thrombopoietin, which have been shown to influence the development of ARDS (Fig. 1) [55, 99, 100].

### The gut–liver–lung axis

The pathogenic mechanisms of ARDS should be considered within a gut–liver–lung axis. Growing evidence indicates that intestinal microbiota and the mucosal immune system of the gut have an important impact on the function of the gastrointestinal tract itself and extra-intestinal organs, such as the lung and the liver [29, 48, 49]. Liver cirrhosis and other liver diseases favor the gut-derived bacterial translocation into the liver and lung by several mechanisms (Fig. 2).

First, patients with liver disorders have intestinal dysbiosis characterized by a significant shift of the microbial composition toward pro-inflammatory bacteria. This gut dysbiosis is accompanied by activation of local intestine immune responses and impaired gut barrier function. A leaky gut barrier facilitates bacterial translocation of live bacteria or their microbial products from the intestinal lumen to the liver, via portal circulation, and to systemic circulation and the lung via the mesenteric lymphatic system [29, 101]. In the lung and the liver, gut microbiota can directly modulate their local immune cells (mainly alveolar macrophages and Kupffer cells, respectively) via activation of toll-like receptors (TLRs) and indirectly via different bacterial metabolites and signaling molecules, such as PAMPs [29, 48, 49, 90]. Activated alveolar macrophages in the lung and Kupffer cells in the liver release pro-inflammatory cytokines, contributing to the initiation and/or progression of lung and liver damage and activation of systemic inflammation [29, 101], which can also cause dysfunction in other organs.

Second, liver dysfunction can imply less capacity of the liver to remove bacteria, bacterial products, and inflammatory mediators from circulation, leading to increased levels of these molecules in blood.

Third, this pathological gut-derived bacterial translocation could cause important changes in the lung microbiome (Fig. 2) [90, 102, 103]. Indeed, pulmonary microbiome is frequently enriched with gut-related bacteria (*Bacteroidetes* and *Enterobacteriaceae*) in critically ill patients [102, 103]. As a consequence of liver diseases, this gut-derived bacteria and accumulation of PAMPs, cytokines and other pro-inflammatory molecules in the systemic circulation can potentially cause or exacerbate lung injury upon TLR-4-mediated activation of intravascular and alveolar macrophages within the lung and recruitment of neutrophils and direct toxic effects of bacterial products on pulmonary microvasculature (Fig. 2) [68, 102–108]. Altogether, the gut–liver–lung axis seems to exert a relevant role in the initiation and modulation of hepatic, pulmonary, and systemic immune responses that contribute to the damage of the liver, the lung, and other organs.



### Extracellular vesicles

Extracellular vesicles (EVs), a term that includes microvesicles (MVs), exosomes, and apoptotic bodies, represent an emerging mechanism of interorgan communication in many diseases, including liver diseases and ARDS [106, 109–111]. Extracellular vesicles are defined as membrane-bound vesicles, ranging 0.1–1.0 µm in diameter, which are released from cells by the budding of the cellular plasma membrane and carrying a diverse cargo, including lipids, proteins, RNAs, and miRNAs. The EVs are recognized as important mediators of interaction between different organs, and they are considered attractive therapeutic targets in different diseases [112, 113]. Notably, the levels of circulating EVs have been reported to be increased both in patients with cirrhosis and ARDS. However, the potential role of circulating EVs in mediating liver–lung communication in the context of ARDS is not currently understood, representing an interesting topic for further investigation.

### Conclusions

Liver injury and hepatotoxicity occur frequently in critically ill patients, and significantly influence their prognosis. Patients with severe hepatic dysfunction are at high risk for irreversible ARDS because of multiple defects in host defense and dysregulation of inflammatory responses. Interrelations between hepatic and pulmonary functions influence the development and progression of ARDS and play a central role in the resolution of lung damage by several mechanisms. First, the liver regulates host defense and modulates systemic inflammation. Also, the liver activates acute inflammatory responses in the lung early in the development of ARDS. Although promoting inflammation can be detrimental in the context of acute lung injury, the liver response to an inflammatory insult is also pro-defense and pro-survival. The understanding of the complex relation between the liver and the lung requires further research in order to improve the clinical management and to identify new diagnostic and therapeutic options for patients with or at risk for ARDS.

### Abbreviations

ARDS: Acute respiratory distress syndrome; DAD: Diffuse alveolar damage; ICU: Intensive care unit; AST: Aspartate aminotransferase; ALT: Alanine aminotransferase; ALP: Alkaline phosphatase; INR: International normalized ratio; PAMPs: Pathogen-associated molecular patterns; TLRs: Toll-like receptors; IL: Interleukin; ENA-78: Epithelial neutrophil-activating peptide-78; TNF: Tumor necrosis factor; MCP-1: Monocyte chemoattractant protein-1; MIP-1α: Macrophage inflammatory protein-1α; PAF: Platelet-activating factor; CCl<sub>4</sub>: Carbon tetrachloride; TBARS: Thiobarbituric acid-reactive substances; APPs: Acute-phase proteins; LPS: Lipopolysaccharides; STAT3: Signal transducer and activator of transcription 3; RelA: v-rel avian reticuloendotheliosis viral oncogene homolog A; NF-κB3: Nuclear factor kappa-light-chain-enhancer of activated B3; SAA: Serum amyloid A; SAP: Serum amyloid P; LBP: Lipopolysaccharide binding protein; CXCL1: Chemokine (C-X-C motif) ligand 1; EVs: Extracellular vesicles; MVs: Microvesicles; RNAs: Ribonucleic acids; miRNAs: Micro-ribonucleic acids

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### Authors' contributions

RH and GS did the design, acquisition of information, and drafting and revising the manuscript. IA, EL, AF, and AdL were responsible for the acquisition of information. JV, LM, RB, and JAL did the revising of the work. The authors read and approved the final manuscript.

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**Competing interests**

The authors declare that they have no competing interests.

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**References**

1. Definition Task Force ARDS, Ranieri VM, Rubenfeld GD, Thompson BT, Ferguson ND, Caldwell E et al (2012) Acute respiratory distress syndrome: the Berlin definition. *JAMA* 307(23):2526–2533
2. Matthay MA, Ware LB, Zimmerman GA (2012) The acute respiratory distress syndrome. *J Clin Invest* 122(8):2731–2740
3. Bellani G, Laffey JG, Pham T, Fan E, Brochard L, Esteban A et al (2016) Epidemiology, patterns of care, and mortality for patients with acute respiratory distress syndrome in intensive care units in 50 countries. *JAMA* 315(8):788–800
4. Matthay MA, Zemans RL (2011) The acute respiratory distress syndrome: pathogenesis and treatment. *Annu Rev Pathol* 6:147–163
5. Matuschak GM, Rinaldo JE (1988) Organ interactions in the adult respiratory distress syndrome during sepsis. Role of the liver in host defense. *Chest* 94(2):400–406
6. Rogers DE (1960) Host mechanisms which act to remove bacteria from the blood stream. *Bacteriol Rev* 24(1):50–66
7. Kolaczowska E, Jenne CN, Surewaard BGJ, Thanabalasuriar A, Lee W-Y, Sanz M-J et al (2015) Molecular mechanisms of NET formation and degradation revealed by intravital imaging in the liver vasculature. *Nat Commun* 6:6673
8. Kaplan JE, Saba TM (1978) Platelet removal from the circulation by the liver and spleen. *Am J Phys* 235(3):H314–H320
9. Bradfield JW (1974) Control of spillover. The importance of Kupffer-cell function in clinical medicine. *Lancet Lond Engl* 2(7885):883–886
10. Guillot A, Tacke F (2019) Liver macrophages: old dogmas and new insights. *Hepatology* 3(6):730–743
11. Khanlou H, Souto H, Lippmann M, Muñoz S, Rothstein K, Ozden Z (1999) Resolution of adult respiratory distress syndrome after recovery from fulminant hepatic failure. *Am J Med Sci* 317(2):134–136
12. Ali M, Wall WJ (1990) Resolution of the adult respiratory distress syndrome following colectomy and liver transplantation. *Chest* 98(4):1032–1034
13. Doyle HR, Marino IR, Miro A, Scott V, Martin M, Fung J et al (1993) Adult respiratory distress syndrome secondary to end-stage liver disease - successful outcome following liver transplantation. *Transplantation* 55(2):292–296
14. Siore AM, Parker RE, Stecenko AA, Cuppels C, McKean M, Christman BW et al (2005) Endotoxin-induced acute lung injury requires interaction with the liver. *Am J Physiol Lung Cell Mol Physiol* 289(5):L769–L776
15. Matuschak GM (1994) Liver-lung interactions in critical illness. *New Horiz* 2(4):488–504
16. Cerra FB, West M, Billiar TR, Holman RT, Simmons R (1989) Hepatic dysfunction in multiple systems organ failure as a manifestation of altered cell-cell interaction. *Prog Clin Biol Res* 308:563–573
17. Matuschak GM (1996) Lung-liver interactions in sepsis and multiple organ failure syndrome. *Clin Chest Med* 17(1):83–98
18. Wang Y, Liu W, Liu X, Sheng M, Pei Y, Lei R et al (2015) Role of liver in modulating the release of inflammatory cytokines involved in lung and multiple organ dysfunction in severe acute pancreatitis. *Cell Biochem Biophys* 71(2):765–776
19. Matuschak GM, Martin DJ (1987) Influence of end-stage liver failure on survival during multiple systems organ failure. *Transplant Proc* 19(4 Suppl 3):40–46
20. Katz S, Grosfeld JL, Gross K, Plager DA, Ross D, Rosenthal RS et al (1984) Impaired bacterial clearance and trapping in obstructive jaundice. *Ann Surg* 199(1):14–20
21. Loegering DJ, Blumenstock FA (1985) Depressing hepatic macrophage complement receptor function causes increased susceptibility to endotoxemia and infection. *Infect Immun* 47(3):659–664
22. DeCamp MM, Warner AE, Molina RM, Brain JD (1992) Hepatic versus pulmonary uptake of particles injected into the portal circulation in sheep. Endotoxin escapes hepatic clearance causing pulmonary inflammation. *Am Rev Respir Dis* 146(1):224–231

23. Hawker F (1991) Liver dysfunction in critical illness. *Anaesth Intensive Care* 19(2):165–181
24. Yang P, Formanek P, Scaglione S, Afshar M (2019) Risk factors and outcomes of acute respiratory distress syndrome in critically ill patients with cirrhosis. *Hepato Res Off J Jpn Soc Hepatol* 49(3):335–343
25. Gacouin A, Locufier M, Uhel F, Letheulle J, Bouju P, Fillatre P et al (2016) Liver cirrhosis is independently associated with 90-day mortality in ARDS patients. *Shock* 45(1):16–21
26. Schwartz DB, Bone RC, Balk RA, Szidon JP (1989) Hepatic dysfunction in the adult respiratory distress syndrome. *Chest* 95(4):871–875
27. Wang CY, Calfee CS, Paul DW, Janz DR, May AK, Zhuo H et al (2014) One-year mortality and predictors of death among hospital survivors of acute respiratory distress syndrome. *Intensive Care Med* 40(3):388–396
28. Dong V, Sun K, Gottfried M, Cardoso FS, McPhail MJ, Stravitz RT et al (2020) Significant lung injury and its prognostic significance in acute liver failure: a cohort analysis. *Liver Int Off J Int Assoc Study Liver* 40(3):654–663
29. Albillas A, Gottardi A, de Rescigno M (2019) The gut-liver axis in liver disease: pathophysiological basis for therapy. *J Hepatol*
30. Matuschak GM, Mattingly ME, Tredway TL, Lechner AJ (1994) Liver-lung interactions during *E. coli* endotoxemia. TNF- $\alpha$ :leukotriene axis. *Am J Respir Crit Care Med* 149(1):41–49
31. Patterson EK, Yao LJ, Ramic N, Lewis JF, Cepinskas G, McCaig L et al (2013) Lung-derived mediators induce cytokine production in downstream organs via an NF- $\kappa$ B-dependent mechanism. *Mediat Inflamm* 2013:586895
32. Karcz M, Bankey B, Schwaiberger D, Lachmann B, Papadakos PJ (2012) Acute respiratory failure complicating advanced liver disease. *Semin Respir Crit Care Med* 33(1):96–110
33. Horvatits T, Drolz A, Trauner M, Fuhrmann V. Liver injury and failure in critical illness. *Hepato* 2019;
34. Kramer L, Jordan B, Druml W, Bauer P, Metnitz PGH, Austrian Epidemiologic Study on Intensive Care, ASDI Study Group et al (2007) *Crit Care Med* 35(4):1099–1104
35. Dizier S, Forel J-M, Ayzac L, Richard J-C, Hraiech S, Lehingue S et al (2015) Early hepatic dysfunction is associated with a worse outcome in patients presenting with acute respiratory distress syndrome: a post-hoc analysis of the ACURASYS and PROSEVA studies. *PLoS One* 10(12):e0144278
36. Lescot T, Karvellas C, Beaussier M, Magder S (2012) Acquired liver injury in the intensive care unit. *Anesthesiology* 117(4): 898–904
37. Marshall JC (2013) The liver in sepsis: shedding light on the cellular basis of hepatocyte dysfunction. *Crit Care Lond Engl* 17(3):153
38. Fuhrmann V, Kneidinger N, Herkner H, Heinz G, Nikfardjam M, Bojic A et al (2011) Impact of hypoxic hepatitis on mortality in the intensive care unit. *Intensive Care Med* 37(8):1302–1310
39. Zhai R, Sheu CC, Su L, Gong MN, Tejera P, Chen F et al (2009) Serum bilirubin levels on ICU admission are associated with ARDS development and mortality in sepsis. *Thorax* 64(9):784–790
40. Kortgen A, Paxian M, Werth M, Recknagel P, Rauchfuss F, Lupp A et al (2009) Prospective assessment of hepatic function and mechanisms of dysfunction in the critically ill. *Shock* 32(4):358–365
41. Sakka SG (2007) Assessing liver function. *Curr Opin Crit Care* 13(2):207–214
42. Vincent JL, Moreno R, Takala J, Willatts S, De Mendonça A, Bruining H et al (1996) The SOFA (sepsis-related organ failure assessment) score to describe organ dysfunction/failure. On behalf of the working group on sepsis-related problems of the European Society of Intensive Care Medicine. *Intensive Care Med* 22(7):707–710
43. Sheu C-C, Gong MN, Zhai R, Chen F, Bajwa EK, Clardy PF et al (2010) Clinical characteristics and outcomes of sepsis-related vs non-sepsis-related ARDS. *Chest* 138(3):559–567
44. Praaning-van Dalen DP, Brouwer A, Knook DL (1981) Clearance capacity of rat liver Kupffer, endothelial, and parenchymal cells. *Gastroenterology* 81(6):1036–1044
45. Freudenberg MA, Freudenberg N, Galanos C (1982) Time course of cellular distribution of endotoxin in liver, lungs and kidneys of rats. *Br J Exp Pathol* 63(1):56–65
46. Bilzer M, Roggel F, Gerbes AL (2006) Role of Kupffer cells in host defense and liver disease. *Liver Int Off J Int Assoc Study Liver* 26(10):1175–1186
47. Triantafyllou E, Woollard KJ, MJW MP, Antoniadis CG, Possamai LA (2018) The role of monocytes and macrophages in acute and acute-on-chronic liver failure. *Front Immunol* 9:2948
48. Young RP, Hopkins RJ, Marsland B (2016) The gut-liver-lung axis, Modulation of the innate immune response and its possible role in chronic obstructive pulmonary disease. *Am J Respir Cell Mol Biol* 54(2):161–169
49. Enaud R, Prevel R, Ciarlo E, Beaufils F, Wieërs G, Guery B, et al. The gut-lung axis in health and respiratory diseases: a place for inter-organ and inter-kingdom crosstalks. *Front Cell Infect Microbiol* [Internet]. 19 de febrero de 2020 [citado 1 de julio de 2020];10. Disponible en: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7042389/>
50. Tarao K, Moroi T, Nagakura Y, Ikeuchi T, Suyama T, Endo O et al (1979) Relationship between endotoxaemia and protein concentration of ascites in cirrhotic patients. *Gut* 20(3):205–210
51. Tilg H, Wilmer A, Vogel W, Herold M, Nölchen B, Judmaier G et al (1992) Serum levels of cytokines in chronic liver diseases. *Gastroenterology* 103(1):264–274
52. Khoruts A, Stahnke L, McClain CJ, Logan G, Allen JI (1991) Circulating tumor necrosis factor, interleukin-1 and interleukin-6 concentrations in chronic alcoholic patients. *Hepato* 13(2):267–276
53. Devière J, Content J, Denys C, Vandenbussche P, Schandene L, Wybran J et al (1990) Excessive in vitro bacterial lipopolysaccharide-induced production of monokines in cirrhosis. *Hepato* 11(4):628–634
54. Torre D, Zeroli C, Giola M, Ferrario G, Fiori GP, Bonetta G et al (1994) Serum levels of interleukin-1 alpha, interleukin-1 beta, interleukin-6, and tumor necrosis factor in patients with acute viral hepatitis. *Clin Infect Dis Off Publ Infect Dis Soc Am* 18(2):194–198
55. Kobayashi K, Horikami D, Omori K, Nakamura T, Yamazaki A, Maeda S et al (2016) Thromboxane A2 exacerbates acute lung injury via promoting edema formation. *Sci Rep* 6:32109
56. Park WY, Goodman RB, Steinberg KP, Ruzinski JT, Radella F, Park DR et al (2001) Cytokine balance in the lungs of patients with acute respiratory distress syndrome. *Am J Respir Crit Care Med* 164(10 Pt 1):1896–1903
57. Goodman RB, Strieter RM, Martin DP, Steinberg KP, Milberg JA, Maunder RJ et al (1996) Inflammatory cytokines in patients with persistence of the acute respiratory distress syndrome. *Am J Respir Crit Care Med* 154(3 Pt 1):602–611

58. Wiedermann FJ, Mayr AJ, Kaneider NC, Fuchs D, Mutz NJ, Schobersberger W (2004) Alveolar granulocyte colony-stimulating factor and alpha-chemokines in relation to serum levels, pulmonary neutrophilia, and severity of lung injury in ARDS. *Chest* 125(1):212–219
59. Frank JA, Matthay MA (2005) Leukotrienes in acute lung injury. *Am J Respir Crit Care Med* 172(3):261–262
60. Rojas M, Woods CR, Mora AL, Xu J, Brigham KL (2005) Endotoxin-induced lung injury in mice: structural, functional, and biochemical responses. *Am J Physiol Lung Cell Mol Physiol* 288(2):L333–L341
61. Panos RJ, Baker SK (1996) Mediators, cytokines, and growth factors in liver-lung interactions. *Clin Chest Med* 17(1):151–169
62. Tran-Thi TA, Weinhold L, Weinstock C, Hoffmann R, Schulze-Specking A, Northoff H et al (1993) Production of tumor necrosis factor-alpha, interleukin-1 and interleukin-6 in the perfused rat liver. *Eur Cytokine Netw* 4(5):363–370
63. Ramadori G, Christ B (1999) Cytokines and the hepatic acute-phase response. *Semin Liver Dis* 19(2):141–155
64. Hermanns MI, Kasper J, Dubruel P, Pohl C, Uboldi C, Vermeersch V et al (2010) An impaired alveolar-capillary barrier in vitro: effect of proinflammatory cytokines and consequences on nanocarrier interaction. *J R Soc Interface* 7(Suppl 1): S41–S54
65. Callery MP, Kamei T, Mangino MJ, Flye MW (1991) Organ interactions in sepsis. Host defense and the hepatic-pulmonary macrophage axis. *Arch Surg Chic Ill* 1960 126(1):28–32
66. Ferrari RS, Tieppo M, da RDP, Forgiarini LA, Dias AS, Marroni NP (2013) Lung and liver changes due to the induction of cirrhosis in two experimental models. *Arq Gastroenterol* 50(3):208–213
67. Salatti Ferrari R, da Rosa DP, Forgiarini LF, Bona S, Dias AS, Marroni NP (2012) Oxidative stress and pulmonary changes in experimental liver cirrhosis. *Oxidative Med Cell Longev* 2012:486190
68. Massey VL, Poole LG, Siow DL, Torres E, Warner NL, Schmidt RH et al (2015) Chronic alcohol exposure enhances lipopolysaccharide-induced lung injury in mice: potential role of systemic tumor necrosis factor-alpha. *Alcohol Clin Exp Res* 39(10):1978–1988
69. Halbertsma FJJ, Vaneker M, Scheffer GJ, van der Hoeven JG. Cytokines and biotrauma in ventilator-induced lung injury: a critical review of the literature. *Neth J Med* 2005;63(10):382–392.
70. Seekamp A, Warren JS, Remick DG, Till GO, Ward PA (1993) Requirements for tumor necrosis factor-alpha and interleukin-1 in limb ischemia/reperfusion injury and associated lung injury. *Am J Pathol* 143(2):453–463
71. Ahuja N, Andres-Hernando A, Altmann C, Bhargava R, Bacalja J, Webb RG et al (2012) Circulating IL-6 mediates lung injury via CXCL1 production after acute kidney injury in mice. *Am J Physiol Renal Physiol* 303(6):F864–F872
72. Zhang H, Neuhöfer P, Song L, Rabe B, Lesina M, Kurkowski MU et al (2013) IL-6 trans-signaling promotes pancreatitis-associated lung injury and lethality. *J Clin Invest* 123(3):1019–1031
73. Quinton LJ, Jones MR, Robson BE, Mizgerd JP (2009) Mechanisms of the hepatic acute-phase response during bacterial pneumonia. *Infect Immun* 77(6):2417–2426
74. Gabay C, Kushner I (1999) Acute-phase proteins and other systemic responses to inflammation. *N Engl J Med* 340(6): 448–454
75. Suffredini AF, Fantuzzi G, Badolato R, Oppenheim JJ, O'Grady NP (1999) New insights into the biology of the acute phase response. *J Clin Immunol* 19(4):203–214
76. Weber M, Lambeck S, Ding N, Henken S, Kohl M, Deigner HP et al (2012) Hepatic induction of cholesterol biosynthesis reflects a remote adaptive response to pneumococcal pneumonia. *FASEB J Off Publ Fed Am Soc Exp Biol* 26(6):2424–2436
77. Gamble L, Bagby GJ, Quinton LJ, Happel KI, Mizgerd JP, Zhang P et al (2009) The systemic and pulmonary LPS binding protein response to intratracheal lipopolysaccharide. *Shock* 31(2):212–217
78. Quinton LJ, Jones MR, Robson BE, Simms BT, Whitsett JA, Mizgerd JP (2008) Alveolar epithelial STAT3, IL-6 family cytokines, and host defense during *Escherichia coli* pneumonia. *Am J Respir Cell Mol Biol* 38(6):699–706
79. Quinton LJ, Blahna MT, Jones MR, Allen E, Ferrari JD, Hilliard KL et al (2012) Hepatocyte-specific mutation of both NF- $\kappa$ B RelA and STAT3 abrogates the acute phase response in mice. *J Clin Invest* 122(5):1758–1763
80. Niemi K, Teirilä L, Lappalainen J, Rajamäki K, Baumann MH, Öörni K et al (2011) Serum amyloid A activates the NLRP3 inflammasome via P2X7 receptor and a cathepsin B-sensitive pathway. *J Immunol* 186(11):6119–6128
81. Shah C, Hari-Dass R, Raynes JG, Serum amyloid A (2006) Is an innate immune opsonin for gram-negative bacteria. *Blood* 108(5):1751–1757
82. Zhang W, Xu W, Xiong S (2011) Macrophage differentiation and polarization via phosphatidylinositol 3-kinase/Akt-ERK signaling pathway conferred by serum amyloid P component. *J Immunol* 187(4):1764–1777
83. Tobias PS, Mathison J, Mintz D, Lee JD, Kravchenko V, Kato K et al (1992) Participation of lipopolysaccharide-binding protein in lipopolysaccharide-dependent macrophage activation. *Am J Respir Cell Mol Biol* 7(3):239–245
84. Mold C, Du Clos TW (2006) C-reactive protein increases cytokine responses to *Streptococcus pneumoniae* through interactions with Fc gamma receptors. *J Immunol* 176(12):7598–7604
85. Barna BP, Deodhar SD, Gautam S, Yen-Lieberman B, Roberts D (1984) Macrophage activation and generation of tumoricidal activity by liposome-associated human C-reactive protein. *Cancer Res* 44(1):305–310
86. Hilliard KL, Allen E, Traber KE, Yamamoto K, Stauffer NM, Wasserman GA et al (2015) The lung-liver axis: a requirement for maximal innate immunity and hepatoprotection during pneumonia. *Am J Respir Cell Mol Biol* 53(3):378–390
87. Frank JA, Wray CM, McAuley DF, Schwendener R, Matthay MA (2006) Alveolar macrophages contribute to alveolar barrier dysfunction in ventilator-induced lung injury. *Am J Physiol Lung Cell Mol Physiol* 291(6):L1191–L1198
88. Renckens R, van Westerloo DJ, Roelofs JJTH, Pater JM, Schultz MJ, Florquin S et al (2008) Acute phase response impairs host defense against *Pseudomonas aeruginosa* pneumonia in mice. *Crit Care Med* 36(2):580–587
89. Renckens R, Roelofs JJTH, Knapp S, de Vos AF, Florquin S, van der Poll T (2006) The acute-phase response and serum amyloid A inhibit the inflammatory response to *Acinetobacter baumannii* pneumonia. *J Infect Dis* 193(2):187–195
90. Sakamori R, Takehara T, Ohnishi C, Tatsumi T, Ohkawa K, Takeda K et al (2007) Signal transducer and activator of transcription 3 signaling within hepatocytes attenuates systemic inflammatory response and lethality in septic mice. *Hepatology* 46(5):1564–1573
91. Postic C, Dentin R, Girard J (2004) Role of the liver in the control of carbohydrate and lipid homeostasis. *Diabetes Metab* 30(5):398–408
92. Hofmann AF (1999) The continuing importance of bile acids in liver and intestinal disease. *Arch Intern Med* 159(22): 2647–2658

93. Rui L (2014) Energy metabolism in the liver. *Compr Physiol* 4(1):177–197
94. Dani C, Martelli E, Tronchin M, Buonocore G, Longini M, Di Filippo A et al (2004) Bilirubin influence on oxidative lung damage and surfactant surface tension properties. *Pediatr Pulmonol* 38(3):179–185
95. Chopra M, Reuben JS, Sharma AC (2009) Acute lung injury: apoptosis and signaling mechanisms. *Exp Biol* 234(4):361–371
96. Rodrigues CMP, Solá S, Brito MA, Brites D, Moura JJG (2002) Bilirubin directly disrupts membrane lipid polarity and fluidity, protein order, and redox status in rat mitochondria. *J Hepatol* 36(3):335–341
97. Cesaratto L, Calligaris SD, Vascotto C, Deganuto M, Bellarosa C, Quadrioglio F et al (2007) Bilirubin-induced cell toxicity involves PTEN activation through an APE1/Ref-1-dependent pathway. *J Mol Med* 85(10):1099–1112
98. Fernandes A, Falcão AS, Silva RFM, Gordo AC, Gama MJ, Brito MA et al (2006) Inflammatory signalling pathways involved in astroglial activation by unconjugated bilirubin. *J Neurochem* 96(6):1667–1679
99. Cuccurullo A, Greco E, Lupia E, De Giuli P, Bosco O, Martin-Conte E et al (2016) Blockade of thrombopoietin reduces organ damage in experimental endotoxemia and polymicrobial sepsis. *PLoS One* 11(3):e0151088
100. Wang Z, Li W, Guo Q, Wang Y, Ma L, Zhang X (2018) Insulin-like growth factor-1 signaling in lung development and inflammatory lung diseases. *Biomed Res Int* 2018:6057589
101. Acharya C, Sahingur SE, Bajaj JS. Microbiota, cirrhosis, and the emerging oral-gut-liver axis. *JCI Insight*. 05 de 2017;2(19).
102. Dickson RP (2018) The lung microbiome and ARDS. It is time to broaden the model. *Am J Respir Crit Care Med* 197(5): 549–551
103. Mukherjee S, Hanidziar D (2018) More of the gut in the lung: how two microbiomes meet in ARDS. *Yale J Biol Med* 91(2):143–149
104. Dickson RP, Singer BH, Newstead MW, Falkowski NR, Erb-Downward JR, Standiford TJ et al (2016) Enrichment of the lung microbiome with gut bacteria in sepsis and the acute respiratory distress syndrome. *Nat Microbiol* 1(10):16113
105. Deitch EA (2010) Gut lymph and lymphatics: a source of factors leading to organ injury and dysfunction. *Ann N Y Acad Sci* 1207(Suppl 1):E103–E111
106. Kojima M, Gimenes-Junior JA, Chan TW, Eliceiri BP, Baird A, Costantini TW et al (2018) Exosomes in postshock mesenteric lymph are key mediators of acute lung injury triggering the macrophage activation via toll-like receptor 4. *FASEB J Off Publ Fed Am Soc Exp Biol* 32(1):97–110
107. de Jong PR, González-Navajas JM, Jansen NJG (2016) The digestive tract as the origin of systemic inflammation. *Crit Care Lond Engl* 20(1):279
108. Massey VL (2015) Potential role of the gut/liver/lung axis in alcohol-induced tissue pathology. *Biomolecules* 5(4):2477–2503
109. Hirsova P, Ibrahim SH, Verma VK, Morton LA, Shah VH, LaRusso NF et al (2016) Extracellular vesicles in liver pathobiology: small particles with big impact. *Hepatol* 64(6):2219–2233
110. Maji S, Matsuda A, Yan IK, Parasramka M, Patel T (2017) Extracellular vesicles in liver diseases. *Am J Physiol Gastrointest Liver Physiol* 312(3):G194–G200
111. Malhi H (2019) Emerging role of extracellular vesicles in liver diseases. *Am J Physiol Gastrointest Liver Physiol* 317(5): G739–G749
112. McVey M, Tabuchi A, Kuebler WM (2012) Microparticles and acute lung injury. *Am J Physiol Lung Cell Mol Physiol* 303(5):L364–L381
113. Rautou P-E, Bresson J, Sainte-Marie Y, Vion A-C, Paradis V, Renard J-M et al (2012) Abnormal plasma microparticles impair vasoconstrictor responses in patients with cirrhosis. *Gastroenterology* 143(1):166–176.e6

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